

Product datasheet

Anti-cetA antibody ab22172

1 Image

Overview

| | |
|----------------------------|---|
| Product name | Anti-cetA antibody |
| Description | Mouse polyclonal to cetA |
| Host species | Mouse |
| Tested applications | Suitable for: WB |
| Species reactivity | Reacts with <i>Campylobacter jejuni</i> . Not yet tested in other species. |
| Immunogen | Fusion protein: SVVKIDHILYKSNMNLNNGAQNFNLESVDPISNLCQDERAQQGVINELSS ETELNLAKEFIKDNAKKAIEESSQDYIDQKAYDAVNDIKSLEQRSAIL , corresponding to amino acids 355/454 of <i>Campylobacter jejuni</i> methyl-accepting chemotaxis protein. |

 [Run BLAST with](#)  [Run BLAST with](#)

General notes Produced from outbred CD1 mice

This antibody was raised by a genetic immunization technique. Genetic immunization can be used to generate antibodies by directly delivering antigen-coding DNA into the animal, rather than injecting a protein or peptide (Tang *et al.* [PubMed: 1545867](#); Chambers and Johnston [PubMed: 12910245](#); Barry and Johnston [PubMed: 9234514](#)). The animal's cells produce the protein, which stimulates the animal's immune system to produce antibodies against that particular protein. A vector coding for a partial fusion protein was used for genetic immunisation of a mouse and the resulting serum was tested in Western blot against an *E.coli* lysate containing that partial fusion protein. Genetic immunization offers enormous advantages over the traditional protein-based immunization method. DNA is faster, cheaper and easier to produce and can be produced by standard techniques readily amenable to automation. Furthermore, the antibodies generated by genetic immunization are usually of superior quality with regard to specificity, affinity and recognizing the native protein.

Properties

| | |
|-----------------------------|--|
| Form | Liquid |
| Storage instructions | Shipped at 4°C. Store at +4°C short term (1-2 weeks). Upon delivery aliquot. Store at -20°C long term. |
| Storage buffer | Constituents: 50% Glycerol |

| | |
|-------------------------------|---|
| Purity | Whole antiserum |
| Primary antibody notes | This antibody was raised by a genetic immunization technique. Genetic immunization can be used to generate antibodies by directly delivering antigen-coding DNA into the animal, rather than injecting a protein or peptide (Tang <i>et al.</i> PubMed: 1545867 ; Chambers and Johnston PubMed: 12910245 ; Barry and Johnston PubMed: 9234514). The animal's cells produce the protein, which stimulates the animal's immune system to produce antibodies against that particular protein. A vector coding for a partial fusion protein was used for genetic immunisation of a mouse and the resulting serum was tested in Western blot against an <i>E.coli</i> lysate containing that partial fusion protein. Genetic immunization offers enormous advantages over the traditional protein-based immunization method. DNA is faster, cheaper and easier to produce and can be produced by standard techniques readily amenable to automation. Furthermore, the antibodies generated by genetic immunization are usually of superior quality with regard to specificity, affinity and recognizing the native protein. |
| Clonality | Polyclonal |
| Isotype | IgG |

Applications

Our [Abpromise guarantee](#) covers the use of **ab22172** in the following tested applications.

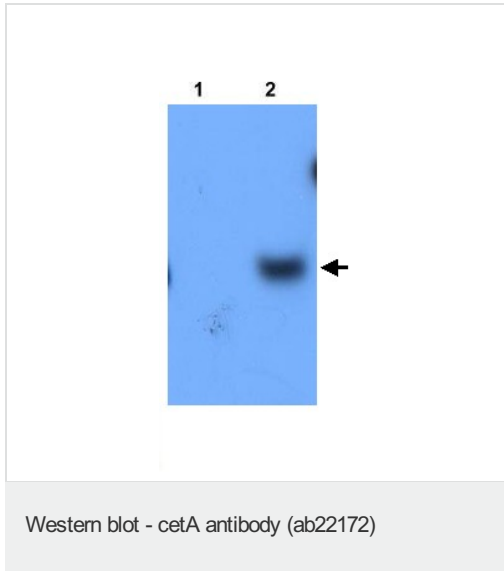
The application notes include recommended starting dilutions; optimal dilutions/concentrations should be determined by the end user.

| Application | Abreviews | Notes |
|-------------|-----------|--|
| WB | | 1/1000. Predicted molecular weight: 51 kDa. This antibody has been tested in Western blot against an <i>E.coli</i> lysate containing the partial recombinant fusion protein used as an immunogen. We have no data on detection of endogenous protein. |

Target

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|------------------|--|
| Relevance | Bacterial chemotactic-signal transducers are proteins that respond to changes in the concentration of attractants and repellents in the environment, and transduce a signal from the outside to the inside of the cell. These proteins undergo two covalent modifications: deamidation and reversible methylation. Attractants increase the level of methylation while repellents decrease it. The methyl groups are added by the methyl-transferase cheR and are removed by the methylesterase cheB |
|------------------|--|

Images



All lanes : Anti-cetA antibody (ab22172) at 1/1000 dilution

Lane 1 : Total protein extract from E. coli with ~50ng to 100ng of a negative control fusion protein with an irrelevant antigen at 20 ug

Lane 2 : Total protein extract from E. coli with ~50ng to 500ng of the antigen fusion protein at 20 ug

Secondary

All lanes : Rabbit anti-mouse IgG + IgM, (H+L) horseradish peroxidase conjugated at 1/5000 dilution

Predicted band size: 51 kDa

The molecular weight of the band on the western blot does not correspond to the predicted band size above (predicted from the molecular weight of the natural protein) because of the additional mass of the fusion and because the fusion protein only contains a partial fragment of the gene.

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