ab108720

Chlamydia trachomatis IgG Human ELISA Kit

Instructions for Use

For the qualitative determination of IgG class antibodies against Chlamydia trachomatis in Human serum or plasma (citrate).

This product is for research use only and is not intended for in vitro diagnostic use.
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1. **Introduction**

ab108720 Chlamydia trachomatis IgG Human ELISA Kit is intended for the qualitative determination of IgG class antibodies against Chlamydia trachomatis in human serum or plasma (citrate).

Chlamydiae are nonmotile, Gram negative and obligatory intracellular growing bacteria which form characteristic inclusions within the cytoplasm of parasitized cells. They are easily visible in the light microscope. Three different Chlamydia species pathogenic for humans are known: Chlamydia trachomatis, Chlamydia pneumoniae and Chlamydia psittaci, and one species only pathogenic for animals (C. pecorum). Chlamydia trachomatis is the most prevalent agent of sexually transmitted diseases worldwide (400-500 million cases) and the number of infections is constantly growing, during childbirth, causing conjunctivitis or pneumonia in newborns. Untreated cases of chlamydial infection can lead to chronic salpingitis, possibly resulting in ectopic pregnancy or infertility. In males, C. trachomatis is a major cause of non-gonococcal urethritis. A severe problem in Chlamydia infections is the frequent asymptomatic insidious course which may result in the initiation of chronic diseases. In many instances primary infections are not recognized and only the sequelae caused by ascended, persisting agents are diagnosed.
2. Assay Summary

*ab108720* is for the qualitative immunoenzymatic determination of IgG-class antibodies against *Chlamydia trachomatis* and is based on the ELISA (Enzyme-linked Immunosorbent Assay) technique.

Microtiter strip wells are precoated with *Chlamydia trachomatis* antigens to bind corresponding antibodies of the specimen.

After washing the wells to remove all unbound sample material horseradish peroxidase (HRP) labelled anti-human IgG conjugate is added. This conjugate binds to the captured *Chlamydia*-specific antibodies.

The immune complex formed by the bound conjugate is visualized by adding Tetramethylbenzidine (TMB) substrate which gives a blue reaction product. The intensity of this product is proportional to the amount of *Chlamydia*-specific IgG antibodies in the specimen.

Sulphuric acid is added to stop the reaction. This produces a yellow endpoint color. Absorption at 450 nm is read using an ELISA microwell plate reader.
3. Kit Contents

- Chlamydia trachomatis Coated Wells (IgG): 12 breakapart 8-well snap-off strips coated with Chlamydia trachomatis antigen; in resealable aluminium foil.
- IgG Sample Diluent ***: 1 bottle containing 100 ml of buffer for sample dilution; pH 7.2 ± 0.2; colored yellow; ready to use; white cap.
- Stop Solution: 1 bottle containing 15 ml sulphuric acid, 0.2 mol/l; ready to use; red cap.
- Washing Solution (20x conc.)*: 1 bottle containing 50 ml of a 20-fold concentrated buffer (pH 7.2 ± 0.2) for washing the wells; white cap.
- Chlamydia trachomatis anti-IgG Conjugate**: 1 bottle containing 20 ml of peroxidase labelled rabbit antibody to human IgG; colored blue, ready to use; black cap.
- TMB Substrate Solution: 1 bottle containing 15 ml 3,3',5,5'-tetramethylbenzidine (TMB); ready to use; yellow cap.
- Chlamydia trachomatis IgG Positive Control***: 1 bottle containing 2 ml; colored yellow; ready to use; red cap.
- Chlamydia trachomatis IgG Cut-off Control***: 1 bottle containing 3 ml; colored yellow; ready to use; green cap.
- Chlamydia trachomatis IgG Negative Control***: 1 bottle containing 2 ml; colored yellow; ready to use; blue cap.
• Strip holder.
• 1 Cover foil.

* contains 0.1 % Bronidox L after dilution
** contains 0.2 % Bronidox L
*** contains 0.1 % Kathon

4. Storage and Handling

The reagents are stable up to the expiry date stated on the label when stored at 2-8 °C.

5. Additional Materials Required

• ELISA microwell plate reader, equipped for the measurement of absorbance at 450/620 nm
• Incubator 37 °C
• Manual or automatic equipment for rinsing wells
• Pipettes to deliver volumes between 10 and 1000 µl
• Vortex tube mixer
• Deionised or (freshly) distilled water
• Disposable tubes
• Timer
6. Preparation of Reagents

1. It is very important to bring all reagents, samples and standards to room temperature (22-28°C) before starting the test run.

2. **Coated snap-off Strips:** The ready to use break apart snap-off strips are coated with Chlamydia trachomatis antigen. Store at 2-8°C. Immediately after removal of strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2-8 °C; stability until expiry date.

3. **Chlamydia trachomatis anti-IgG Conjugate:** The bottle contains 20 ml of a solution with anti-human-IgG horseradish peroxidase, buffer, stabilizers, preservatives and an inert blue dye. The solution is ready to use. Store at 2-8°C. After first opening stability until expiry date when stored at 2-8 °C.

4. **Controls:** The bottles labelled with Positive, Cut-off and Negative Control contain a ready to use control solution. It contains 0.1% Kathon and has to be stored at 2-8°C. After first opening stability until expiry date when stored at 2-8 °C.

5. **IgG Sample Diluent:** The bottle contains 100 ml phosphate buffer, stabilizers, preservatives and an inert yellow dye. It is used for the dilution of the specimen. This ready to use solution has to be stored at 2-8°C. After first opening stability until expiry date when stored at 2-8 °C.
6. **Washing Solution (20x conc.):** The bottle contains 50 ml of a concentrated buffer, detergents and preservatives. Dilute Washing Solution 1+19; e.g. 10 ml Washing Solution + 190 ml fresh and germ free redistilled water. The diluted buffer is stable for 5 days at room temperature. *Crystals in the solution disappear by warming up to 37°C in a water bath. After first opening stability until expiry date when stored at 2-8°C.*

7. **TMB Substrate Solution:** The bottle contains 15 ml of a tetramethylbenzidine/hydrogen peroxide system. The reagent is ready to use and has to be stored at 2-8°C away from the light. *The solution should be colorless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away. After first opening stability until expiry date when stored at 2-8°C.*

8. **Stop Solution:** The bottle contains 15 ml 0.2 M sulphuric acid solution. This ready to use solution has to be stored at 2-8°C. *After first opening stability until expiry date.*

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7. **Preparation and Collection of Specimen**

Use human serum or plasma (citrate) samples with this assay. If the assay is performed within 5 days after sample collection, the specimen should be kept at 2-8°C; otherwise they should be aliquoted and stored deep-frozen (-20 to -70°C). If samples are
stored frozen, mix thawed samples well before testing. *Avoid repeated freezing and thawing.*

*Heat inactivation of samples is not recommended.*

**Sample Dilution:**

Before assaying, all samples should be diluted 1+100 with IgG Sample Diluent. Dispense 10µl sample and 1ml IgG Sample Diluent into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

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**8. Assay Method**

**Test Preparation**

Please read the test protocol carefully **before** performing the assay. Result reliability depends on strict adherence to the test protocol as described. If performing the test on ELISA automatic systems we recommend to increase the washing steps from three to five and the volume of washing solution from 300µl to 350µl to avoid washing effects. Prior to commencing the assay, the distribution and identification plan for all specimens and controls should be carefully
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established. Select the required number of microtiter strips or wells and insert them into the holder. Please allocate at least:

- 1 well (e.g. A1) for the blank
- 1 well (e.g. B1) for the negative control
- 2 wells (e.g. C1+D1) for the cut-off control and
- 1 well (e.g. E1) for the positive control

- It is left to the user to determine controls and samples in duplicate, if necessary.
- Perform all assay steps in the order given and without any appreciable delays between the steps.
- A clean, disposable tip should be used for dispensing each standard and each sample.
- Adjust the incubator to 37°C ± 1°C.

Assay Procedure:

1. Dispense 100 µl controls and diluted samples into their respective wells. Leave well A1 for substrate blank.
2. Cover wells with the foil supplied in the kit.
3. **Incubate for 1 hour ± 5 min at 37°C.**
4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300µl of Washing Solution. Avoid overflows from the reaction wells. The soak time between each wash cycle should be >5sec.
At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!

*Note: Washing is critical! Insufficient washing results in poor precision and falsely elevated absorbance values.*

5. Dispense 100µl Chlamydia trachomatis anti-IgG Conjugate into all wells except for the blank well (e.g. A1). Cover with foil.

6. **Incubate for 30 min at room temperature.** *Do not expose to direct sunlight.*

7. Repeat step 4.

8. Dispense 100µl TMB Substrate Solution into all wells.

9. **Incubate for exactly 15 min at room temperature in the dark.**

10. Dispense 100µl Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution.

   *Any blue colour developed during the incubation turns into yellow.*

   *Note: Highly positive samples can cause dark precipitates of the chromogen! These precipitates have an influence when reading the optical density. Predilution of the sample with physiological sodium chloride solution, for example 1+1, is recommended. Then dilute the sample 1+100 with dilution buffer and multiply the results in NTU by 2.*

11. Measure the absorbance of the specimen at 450/620 nm within 30 min after addition of the Stop Solution.
Measurement:

Adjust the ELISA Microwell Plate Reader to zero using the standard 0.

*If - due to technical reasons - the ELISA reader cannot be adjusted to zero using the substrate blank in well A1, subtract the absorbance value of well A1 from all other absorbance values measured in order to obtain reliable results!*

Measure the absorbance of all wells at 450 nm and record the absorbance values for each standard and sample in the distribution and identification plan.

Dual wavelength reading using 620 nm as reference wavelength is recommended.

Where applicable calculate the mean absorbance values of all duplicates.
9. Data Analysis

A. Run Validation Criteria

In order for an assay to be considered valid, the following criteria must be met:

- **Substrate blank** in A1: Absorbance value $< 0.100$.
- **Negative control** in B1: Absorbance value $< 0.200$ and $< \text{cut-off}$.
- **Cut-off control** in C1 and D1: Absorbance value $0.150 - 1.300$.
- **Positive control** in E1: Absorbance value $> \text{cut-off}$.

If these criteria are not met, the test is not valid and must be repeated.

B. Calculation of Results

The cut-off is the mean absorbance value of the Cut-off control determinations.

*Example: Absorbance value Cut-off control 0.39 + absorbance value Cut-off control 0.37 =0.76 / 2 = 0.38

Cut-off = 0.38
C. Interpretation of Results

Samples are considered **POSITIVE** if the absorbance value is higher than 10% over the cut-off.

Samples with an absorbance value of 10% above or below the cut-off should not be considered as clearly positive or negative.

→ grey zone

It is recommended to repeat the test again 2 - 4 weeks later with a fresh sample. If results in the second test are again in the grey zone the sample has to be considered **NEGATIVE**.

Samples are considered **NEGATIVE** if the absorbance value is lower than 10% below the cut-off.

D. Results in Abcam Units

\[
\text{Sample (mean) absorbance value } \times 10 = \text{[Abcam Units = NTU]} \\
\text{Cut off}
\]

Example: \( \frac{1.204 \times 10}{0.38} = 32 \text{ NTU [Abcam Units = NTU]} \)

- Cut-off: 10 NTU
- Greyzone: 9-11 NTU
- Negative: <9 NTU
- Positive: >11 NTU
E. **Precision**

<table>
<thead>
<tr>
<th>Inter-assay</th>
<th>n</th>
<th>Mean (NTU)</th>
<th>Cv (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pos. Serum</td>
<td>12</td>
<td>61</td>
<td>2.8</td>
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<table>
<thead>
<tr>
<th>Intra-assay</th>
<th>n</th>
<th>Mean (E)</th>
<th>Cv (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pos. Serum</td>
<td>24</td>
<td>2.07</td>
<td>3.7</td>
</tr>
</tbody>
</table>

F. **Diagnostic Specificity**

The diagnostic specificity is defined as the probability of the assay of scoring negative in the absence of the specific analyte. It is 91.3%.

G. **Diagnostic Sensitivity**

The diagnostic sensitivity is defined as the probability of the assay of scoring positive in the presence of the specific analyte. It is > 95%.
H. Interferences

Interferences with hemolytic, lipemic or icteric sera are not observed up to a concentration of 10 mg/ml hemoglobin, 5 mg/ml triglycerides and 0.2 mg/ml bilirubin.

Note: The results refer to the groups of samples investigated; these are not guaranteed specifications.

10. Limitations

- Bacterial contamination or repeated freeze-thaw cycles of the specimen may affect the absorbance values.
- Diagnosis of an infectious disease should not be established on the basis of a single test result. A precise diagnosis should take into consideration clinical history, symptomatology as well as serological data.
- In immunocompromised patients and newborns serological data only have restricted value.
- The Chlamydia trachomatis antigen which is coated on the plates is comprised of elementary bodies. A cross reaction with Chlamydia pneumoniae cannot be excluded with sera containing antibodies to LPS and MOMP.
# 11. Troubleshooting

<table>
<thead>
<tr>
<th>Problem</th>
<th>Cause</th>
<th>Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td>Low signal</td>
<td>Incubation time too short</td>
<td>Try overnight incubation at 4 °C</td>
</tr>
<tr>
<td></td>
<td>Precipitate can form in wells upon substrate addition when concentration of target is too high</td>
<td>Increase dilution factor of sample</td>
</tr>
<tr>
<td></td>
<td>Using incompatible sample type (e.g. serum vs. cell extract)</td>
<td>Detection may be reduced or absent in untested sample types</td>
</tr>
<tr>
<td></td>
<td>Sample prepared incorrectly</td>
<td>Ensure proper sample preparation/dilution</td>
</tr>
<tr>
<td>Large CV</td>
<td>Bubbles in wells</td>
<td>Ensure no bubbles present prior to reading plate</td>
</tr>
<tr>
<td></td>
<td>All wells not washed equally/thoroughly</td>
<td>Check that all ports of plate washer are unobstructed/wash wells as recommended</td>
</tr>
<tr>
<td></td>
<td>Incomplete reagent mixing</td>
<td>Ensure all reagents/master mixes are mixed thoroughly</td>
</tr>
<tr>
<td></td>
<td>Inconsistent pipetting</td>
<td>Use calibrated pipettes &amp; ensure accurate pipetting</td>
</tr>
<tr>
<td></td>
<td>Inconsistent sample preparation or storage</td>
<td>Ensure consistent sample preparation and optimal sample storage conditions (e.g. minimize freeze/thaw cycles)</td>
</tr>
<tr>
<td>High background</td>
<td>Wells are insufficiently washed</td>
<td>Wash wells as per protocol recommendations</td>
</tr>
<tr>
<td>-----------------</td>
<td>---------------------------------</td>
<td>---------------------------------------------</td>
</tr>
<tr>
<td>Contaminated wash buffer</td>
<td>Make fresh wash buffer</td>
<td></td>
</tr>
<tr>
<td>Waiting too long to read plate after adding STOP solution</td>
<td>Read plate immediately after adding STOP solution</td>
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</table>

<table>
<thead>
<tr>
<th>Low sensitivity</th>
<th>Improper storage of ELISA kit</th>
<th>Store all reagents as recommended. Please note all reagents may not have identical storage requirements.</th>
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<td>Using incompatible sample type (e.g. Serum vs. cell extract)</td>
<td>Detection may be reduced or absent in untested sample types</td>
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</table>

For further technical questions please do not hesitate to contact us by email ([technical@abcam.com](mailto:technical@abcam.com)) or phone (select “contact us” on [www.abcam.com](http://www.abcam.com) for the phone number for your region).
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<thead>
<tr>
<th><strong>Abcam in the USA</strong></th>
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<th><strong>Abcam in Europe</strong></th>
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<tbody>
<tr>
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<td>1 Science Park West Avenue</td>
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<td>UK</td>
<td>Hong Kong Science Park</td>
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