ab150686

Trichrome Stain
(Connective Tissue Stain)

Instructions for Use

For the Histological Visualization of Collagenous Connective Tissue Fibers in Tissue Sections

This product is for research use only and is not intended for diagnostic use.

Version 2 Last Updated 31 October 2016
Table of Contents

1. Introduction 3
2. Kit Contents 4
3. Storage and Handling 4
4. Additional Materials Required 5
5. Staining Protocol 5
6. Reference Image 7
1. Introduction

The Trichrome Stain (Connective Tissue Stain) is intended for use in the histological visualization of collagenous connective tissue fibers in tissue sections. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Staining Interpretation:

<table>
<thead>
<tr>
<th>Collagen</th>
<th>Blue</th>
</tr>
</thead>
<tbody>
<tr>
<td>Muscle Fiber</td>
<td>Red</td>
</tr>
<tr>
<td>Nuclei</td>
<td>Black/Blue</td>
</tr>
</tbody>
</table>

Control Tissue:

Lung, Uterus, Small Intestine, Stomach
## 2. Kit Contents

<table>
<thead>
<tr>
<th>Components</th>
<th>Amount</th>
<th>Storage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bouin's Fluid</td>
<td>125 ml</td>
<td>RT</td>
</tr>
<tr>
<td>Weigert’s Iron, Hematoxylin (A)</td>
<td>125 ml</td>
<td>RT</td>
</tr>
<tr>
<td>Weigert’s Iron, Hematoxylin (B)</td>
<td>125 ml</td>
<td>RT</td>
</tr>
<tr>
<td>Biebrich Scarlet/Acid Fuchsin Solution</td>
<td>125 ml</td>
<td>RT</td>
</tr>
<tr>
<td>Phosphomolybdic/Phosphotungstic Acid</td>
<td>125 ml</td>
<td>RT</td>
</tr>
<tr>
<td>Aniline Blue Solution</td>
<td>125 ml</td>
<td>RT</td>
</tr>
<tr>
<td>Acetic Acid Solution (1%)</td>
<td>125 ml</td>
<td>RT</td>
</tr>
</tbody>
</table>

## 3. Storage and Handling

For storage temperatures please see the Table

Keep away from open flame. Refer to the Safety Data Sheet.
4. Additional Materials Required

Absolute Alcohol

Xylene or Xylene Substitute

5. Staining Protocol

1. Deparaffinize sections if necessary and hydrate to distilled water.

2. Preheat Bouin’s Fluid in a water bath to 56-64°C in a fume hood or very well ventilated area.

3. Place slide in preheated Bouin’s Fluid for 60 minutes followed by a 10 minute cooling period.

4. Rinse slide in tap water until section is completely clear.

5. Rinse once in distilled water.

6. Mix equal parts of Weigert’s (A) and Weigert’s (B) and stain slide with working Weigert’s Iron Hematoxylin for 5 minutes.

7. Rinse slide in running tap water for 2 minutes.

8. Apply Biebrich Scarlet / Acid Fuchsin Solution to slide for 15 minutes.
9. Rinse slide in distilled water.

10. Differentiate in Phosphomolybdic/Phosphotungstic Acid Solution for 10-15 minutes or until collagen is not red.

11. Without rinsing, apply Aniline Blue Solution to slide for 5-10 minutes.

12. Rinse slide in distilled water.

13. Apply Acetic Acid Solution (1%) to slide for 3-5 minutes.

14. Dehydrate very quickly in 2 changes of 95% Alcohol, followed by 2 changes of Absolute Alcohol.

15. Clear in Xylene or Xylene Substitute and mount in synthetic resin.
6. Reference Image

Figure 1. Staining using ab150686 - Trichrome Stain Kit (Modified Masson's)

For further technical questions please do not hesitate to contact us by email (technical@abcam.com) or phone (select “contact us” on www.abcam.com for the phone number for your region).