

Product datasheet

Human ATG4D knockout HeLa cell line ab265980

1 Image

Overview

Product name Human ATG4D knockout HeLa cell line

Parental Cell Line HeLa
Organism Human

Mutation description Knockout achieved by using CRISPR/Cas9, Homozygous: 1 bp insertion in exon 1

Passage number <20

Knockout validation Sanger Sequencing

Biosafety level 2

General notesRecommended control: Human wild-type HeLa cell line (ab255928). Please note a wild-type cell line is not automatically included with a knockout cell line order, if required please add recommended wild-type cell line at no additional cost using the code WILDTYPE-TMTK1.

Cryopreservation cell medium: Cell Freezing Medium-DMSO Serum free media, contains 8.7% DMSO in MEM supplemented with methyl cellulose.

Culture medium: DMEM (High Glucose) + 10% FBS

Initial handling guidelines: Upon arrival, the vial should be stored in liquid nitrogen vapor phase and not at -80°C. Storage at -80°C may result in loss of viability.

- 1. Thaw the vial in 37°C water bath approximately 1-2 minutes.
- 2. Transfer the cell suspension (0.8 ml) to a 15 ml/50 ml conical sterile polypropylene centrifuge tube containing 8.4 ml pre-warmed **culture medium**, wash vial with an additional 0.8 ml **culture medium** (total volume 10 ml) to collect remaining cells, and centrifuge at 201 x g (rcf) for 5 minutes at room temperature. 10 ml represents minimum recommended dilution. 20 ml represents maximum recommended dilution.
- 3. Resuspend the cell pellet in 5 ml pre-warmed **culture medium** and count using a haemocytometer (Click here to view haemocytometer protocol) or alternative cell counting method. Based on cell count, seed cells in an appropriate cell culture flask at a density of $2x10^4$ cells/cm². This should allow for confluency within 48 hours. Seeding density is given as a guide only and should be scaled to align with individual lab schedules.
- 4. Incubate the culture at 37°C incubator with 5% CO₂. Cultures should be monitored daily.

Subculture guidelines:

All seeding densities should be based on cell counts gained by established methods. A guide seeding density of 2x10⁴ cells/cm² is recommended for confluency (80-90% confluence) within 48 hours.

A partial media change 24 hours prior to subculture may be helpful to encourage growth, if

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required.

Cells should be passaged when they have achieved 80-90% confluence.

Click here to view the Mammalian cell tissue culture protocol

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Properties

Number of cells 1 x 10⁶ cells/vial, 1 mL

Viability ~90%

Adherent /Suspension Adherent
Tissue Cervix
Cell type epithelial

Disease Adenocarcinoma

Gender Female

STR Analysis Amelogenin X D5S818: 11, 12 D13S317: 12, 13.3 D7S820: 8, 12 D16S539: 9, 10 vWA: 16, 18

TH01: 7 TPOX: 8,12 CSF1PO: 9, 10

Mycoplasma free Yes

Storage instructions Shipped on Dry Ice. Store in liquid nitrogen.

Storage buffer Constituents: 8.7% DMSO, 2% Cellulose, methyl ether

Purity Immunogen affinity purified

Target

Function Cysteine protease required for autophagy, which cleaves the C-terminal part of either MAP1LC3,

GABARAPL2 or GABARAP, allowing the liberation of form I. A subpopulation of form I is subsequently converted to a smaller form (form II). Form II, with a revealed C-terminal glycine, is considered to be the phosphatidylethanolamine (PE)-conjugated form, and has the capacity for

the binding to autophagosomes.

Tissue specificity Mainly expressed in skeletal muscle and, to a lower extent, in testis.

Sequence similarities Belongs to the peptidase C54 family.

Cellular localization Cytoplasm.

Images

Mut CATGAACTCAGTGTCGCCGGCCGCCGCAAGTACCGGAGCAGCAGCCCGGAGGACGCGC

Homozygous: 1 bp insertion in exon 1.

Sanger Sequencing - Human ATG4D knockout HeLa

cell line (ab265980)

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