

# Product datasheet

# Human CEP57 knockout HeLa cell line ab266056

## 3 Images

#### Overview

Product name Human CEP57 knockout HeLa cell line

Parental Cell Line HeLa
Organism Human

Mutation description Knockout achieved by using CRISPR/Cas9, 50 bp deletion in exon 1 and Insertion of the selection

cassette in exon 1

Passage number <20

Knockout validation Sanger Sequencing

Biosafety level

General notes Recommended control: Human wild-type HeLa cell line (ab255928). Please note a wild-type

cell line is not automatically included with a knockout cell line order, if required please add recommended wild-type cell line at no additional cost using the code WILDTYPE-TMTK1.

Cryopreservation cell medium: Cell Freezing Medium-DMSO Serum free media, contains

8.7% DMSO in MEM supplemented with methyl cellulose.

Culture medium: DMEM (High Glucose) + 10% FBS

**Initial handling guidelines:** Upon arrival, the vial should be stored in liquid nitrogen vapor phase and not at -80°C. Storage at -80°C may result in loss of viability.

- 1. Thaw the vial in 37°C water bath approximately 1-2 minutes.
- 2. Transfer the cell suspension (0.8 ml) to a 15 ml/50 ml conical sterile polypropylene centrifuge tube containing 8.4 ml pre-warmed **culture medium**, wash vial with an additional 0.8 ml **culture medium** (total volume 10 ml) to collect remaining cells, and centrifuge at 201 x g (rcf) for 5 minutes at room temperature. 10 ml represents minimum recommended dilution. 20 ml represents maximum recommended dilution.
- 3. Resuspend the cell pellet in 5 ml pre-warmed **culture medium** and count using a haemocytometer (Click here to view haemocytometer protocol) or alternative cell counting method. Based on cell count, seed cells in an appropriate cell culture flask at a density of  $2x10^4$  cells/cm<sup>2</sup>. This should allow for confluency within 48 hours. Seeding density is given as a guide only and should be scaled to align with individual lab schedules.
- 4. Incubate the culture at 37°C incubator with 5% CO<sub>2</sub>. Cultures should be monitored daily.

### Subculture guidelines:

All seeding densities should be based on cell counts gained by established methods.

A guide seeding density of 2x10<sup>4</sup> cells/cm<sup>2</sup> is recommended for confluency (80-90% confluence) within 48 hours.

A partial media change 24 hours prior to subculture may be helpful to encourage growth, if

Cells should be passaged when they have achieved 80-90% confluence.

Click here to view the Mammalian cell tissue culture protocol

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#### **Properties**

**Number of cells** 1 x 10<sup>6</sup> cells/vial, 1 mL

Viability ~90%

Adherent /Suspension Adherent

Tissue Cervix
Cell type epithelial

**Disease** Adenocarcinoma

**Gender** Female

**STR Analysis** Amelogenin X D5S818: 11, 12 D13S317: 12, 13.3 D7S820: 8, 12 D16S539: 9, 10 vWA: 16, 18

TH01: 7 TPOX: 8,12 CSF1PO: 9, 10

Mycoplasma free Yes

**Storage instructions** Shipped on Dry Ice. Store in liquid nitrogen.

Storage buffer Constituents: 8.7% DMSO, 2% Cellulose, methyl ether

**Purity** Immunogen affinity purified

**Target** 

**Function** Centrosomal protein which may be required for microtubule attachment to centrosomes. May act

by forming ring-like structures around microtubules. Mediates nuclear translocation and mitogenic

activity of the internalized growth factor FGF2.

Tissue specificity Ubiquitous.

Sequence similarities Belongs to the translokin family.

**Domain** The C-terminal region mediates the interaction with microtubules and is able to nucleate and

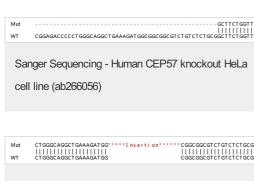
bundles microtubules in vitro.

The centrosome localization domain (CLD) region mediates the localization to centrosomes and

homooligomerization.

**Cellular localization** Nucleus. Cytoplasm. Cytoplasm > cytoskeleton > centrosome.

**Images** 



Allele-1: 50 bp deletion in exon1



Allele-2: Insertion of the selection cassette in exon 1.



Allele-3: Insertion of the selection cassette in exon 1.

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