# abcam

### **Product datasheet**

## Human CSF1R (CSF-1-R) knockout HeLa cell line ab267226

#### 1 Image

Overview

Product name	Human CSF1R (CSF-1-R) knockout HeLa cell line				
Parental Cell Line	HeLa				
Organism	Human				
Mutation description	Knockout achieved by using CRISPR/Cas9, Homozygous: 1 bp deletion in exon 4				
Passage number	<20				
Knockout validation	Sanger Sequencing				
Biosafety level	2				
General notes	<b>Recommended control:</b> Human wild-type HeLa cell line ( <u>ab255928</u> ). Please note a wild-type cell line is not automatically included with a knockout cell line order, if required please add recommended wild-type cell line at no additional cost using the code WILDTYPE-TMTK1.				
	<b>Cryopreservation cell medium:</b> Cell Freezing Medium-DMSO Serum free media, contains 8.7% DMSO in MEM supplemented with methyl cellulose.				
	Culture medium: DMEM (High Glucose) + 10% FBS				
	<b>Initial handling guidelines:</b> Upon arrival, the vial should be stored in liquid nitrogen vapor phase and not at -80°C. Storage at -80°C may result in loss of viability.				
	<ol> <li>Thaw the vial in 37°C water bath for approximately 1-2 minutes.</li> <li>Transfer the cell suspension (0.8 mL) to a 15 mL/50 mL conical sterile polypropylene centrifuge tube containing 8.4 mL pre-warmed culture medium, wash vial with an additional 0.8 mL culture medium (total volume 10 mL) to collect remaining cells, and centrifuge at 201 x g (rcf) for 5 minutes at room temperature. 10 mL represents minimum recommended dilution. 20 mL represents maximum recommended dilution.</li> <li>Resuspend the cell pellet in 5 mL pre-warmed culture medium and count using a haemocytometer or alternative cell counting method. Based on cell count, seed cells in an appropriate cell culture flask at a density of 2x10<sup>4</sup> cells/cm<sup>2</sup>. Seeding density is given as a guide</li> </ol>				
	only and should be scaled to align with individual lab schedules. 4. Incubate the culture at 37°C incubator with 5% CO <sub>2</sub> . Cultures should be monitored daily.				
	<b>Subculture guidelines:</b> All seeding densities should be based on cell counts gained by established methods. A guide seeding density of 2x10 <sup>4</sup> cells/cm <sup>2</sup> is recommended. A partial media change 24 hours prior to subculture may be helpful to encourage growth, if				

required.

Cells should be passaged when they have achieved 80-90% confluence. This product is subject to limited use licenses from The Broad Institute and ERS Genomics Limited, and is developed with patented technology. For full details of the limited use licenses and relevant patents please refer to our **limited use license** and **patent pages**.

We will provide viable cells that proliferate on revival.

#### Properties

Number of cells	1 x 10 <sup>6</sup> cells/vial, 1 mL
Adherent /Suspension	Adherent
Tissue	Cervix
Cell type	epithelial
Disease	Adenocarcinoma
Gender	Female
STR Analysis	Amelogenin X D5S818: 11, 12 D13S317: 12, 13.3 D7S820: 8, 12 D16S539: 9, 10 vWA: 16, 18 TH01: 7 TPOX: 8, 12 CSF1PO: 9, 10
Antibiotic resistance	Puromycin 1.00µg/ml
Mycoplasma free	Yes
Storage instructions	Shipped on Dry Ice. Store in liquid nitrogen.
Storage buffer	Constituents: 8.7% Dimethylsulfoxide, 2% Cellulose, methyl ether

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Function	Protein tyrosine-kinase transmembrane receptor for CSF1 and IL34.			
Tissue specificity	Expressed in bone marrow and in differentiated blood mononuclear cells.			
Sequence similarities	Belongs to the protein kinase superfamily. Tyr protein kinase family. CSF-1/PDGF receptor subfamily. Contains 5 lg-like C2-type (immunoglobulin-like) domains. Contains 1 protein kinase domain.			
Cellular localization	Membrane.			

#### Images



Sanger Sequencing - Human CSF1R knockout HeLa cell line (ab267226)

Homozygous: 1 bp deletion in exon4

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