abcam

Product datasheet

Human FTL knockout HeLa cell line ab265534

4 Images

Overview

Product name Human FTL knockout HeLa cell line

Parental Cell Line HeLa
Organism Human

Mutation description Knockout achieved by using CRISPR/Cas9, 1 bp insertion in exon 1 and 2 bp insertion in exon 1

Passage number <20

Knockout validation Sanger Sequencing, Western Blot (WB)

Tested applications Suitable for: WB

Biosafety level

General notesRecommended control: Human wild-type HeLa cell line (<u>ab255928</u>). Please note a wild-type cell line is not automatically included with a knockout cell line order, if required please add recommended wild-type cell line at no additional cost using the code WILDTYPE-TMTK1.

Cryopreservation cell medium: Cell Freezing Medium-DMSO Serum free media, contains 8.7% DMSO in MEM supplemented with methyl cellulose.

Culture medium: DMEM (High Glucose) + 10% FBS

Initial handling guidelines: Upon arrival, the vial should be stored in liquid nitrogen vapor phase and not at -80°C. Storage at -80°C may result in loss of viability.

- 1. Thaw the vial in 37°C water bath for approximately 1-2 minutes.
- 2. Transfer the cell suspension (0.8 mL) to a 15 mL/50 mL conical sterile polypropylene centrifuge tube containing 8.4 mL pre-warmed culture medium, wash vial with an additional 0.8 mL culture medium (total volume 10 mL) to collect remaining cells, and centrifuge at 201 x g (rcf) for 5 minutes at room temperature. 10 mL represents minimum recommended dilution. 20 mL represents maximum recommended dilution.
- 3. Resuspend the cell pellet in 5 mL pre-warmed culture medium and count using a haemocytometer or alternative cell counting method. Based on cell count, seed cells in an appropriate cell culture flask at a density of 2x10⁴ cells/cm². Seeding density is given as a guide only and should be scaled to align with individual lab schedules.
- 4. Incubate the culture at 37°C incubator with 5% CO₂. Cultures should be monitored daily.

Subculture guidelines:

All seeding densities should be based on cell counts gained by established methods. A guide seeding density of $2x10^4$ cells/cm² is recommended.

A partial media change 24 hours prior to subculture may be helpful to encourage growth, if required.

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Cells should be passaged when they have achieved 80-90% confluence.

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We will provide viable cells that proliferate on revival.

Properties

Number of cells 1 x 10⁶ cells/vial, 1 mL

Adherent /Suspension Adherent
Tissue Cervix
Cell type epithelial

Disease Adenocarcinoma

Gender Female

STR Analysis Amelogenin X D5S818: 11, 12 D13S317: 12, 13.3 D7S820: 8, 12 D16S539: 9, 10 vWA: 16, 18

TH01: 7 TPOX: 8,12 CSF1PO: 9, 10

Antibiotic resistance Puromycin 1.00µg/ml

Mycoplasma free Yes

Storage instructions Shipped on Dry Ice. Store in liquid nitrogen.

Storage buffer Constituents: 8.7% Dimethylsulfoxide, 2% Cellulose, methyl ether

Target

Function Stores iron in a soluble, non-toxic, readily available form. Important for iron homeostasis. Iron is

taken up in the ferrous form and deposited as ferric hydroxides after oxidation. Also plays a role in

delivery of iron to cells. Mediates iron uptake in capsule cells of the developing kidney.

Involvement in diseaseDefects in FTL are the cause of hereditary hyperferritinemia-cataract syndrome (HHCS)

[MIM:600886]. It is an autosomal dominant disease characterized by early-onset bilateral

cataract. Affected patients have elevated level of circulating ferritin. HHCS is caused by mutations

in the iron responsive element (IRE) of the FTL gene.

Defects in FTL are the cause of neurodegeneration with brain iron accumulation type 3 (NBIA3) [MIM:606159]; also known as adult-onset basal ganglia disease. It is a movement disorder with heterogeneous presentations starting in the fourth to sixth decade. It is characterized by a variety of neurological signs including parkinsonism, ataxia, corticospinal signs, mild nonprogressive cognitive deficit and episodic psychosis. It is linked with decreased serum ferritin levels.

Sequence similarities Belongs to the ferritin family.

Contains 1 ferritin-like diiron domain.

Applications

The Abpromise guarantee Our Abpromise guarantee covers the use of ab265534 in the following tested applications.

The application notes include recommended starting dilutions; optimal dilutions/concentrations should be determined by the end user.

Application	Abreviews	Notes
WB		Use at an assay dependent concentration. Predicted molecular weight: 20 kDa.

Images



Western blot - Human FTL knockout HeLa cell line (ab265534)

All lanes : Anti-Ferritin Light Chain antibody [FTL/1386] (ab218400) at 1/1000 dilution

Lane 1 : Wild-type HeLa cell lysate

Lane 2 : FTL knockout HeLa cell lysate

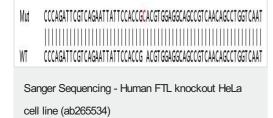
Lysates/proteins at 20 µg per lane.

Performed under reducing conditions.

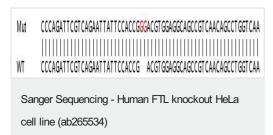
Predicted band size: 20 kDa Observed band size: 20 kDa

Lanes 1-2: Merged signal (red and green). Green - <u>ab218400</u> observed at 20 kDa. Red - Anti-alpha Tubulin antibody [EP1332Y] - Microtubule Marker (<u>ab52866</u>) observed at 50 kDa.

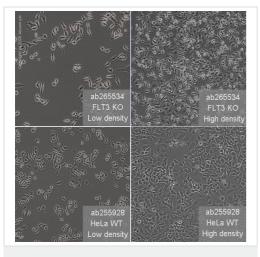
ab218400 was shown to react with Ferritin Light Chain in wild-type HeLa cells in western blot. Loss of signal was observed when knockout cell line ab265534 (knockout cell lysate ab256927) was used. Wild-type HeLa and Ferritin Light Chain knockout HeLa cell lysates were subjected to SDS-PAGE. Membrane was blocked for 1 hour at room temperature in 0.1% TBST with 3% non-fat dried milk. ab218400 and Anti-alpha Tubulin antibody [EP1332Y] - Microtubule Marker (ab52866) overnight at 4°C at a 1 in 1000 dilution and a 1 in 20000 dilution respectively. Blots were developed with Goat anti-Mouse IgG H&L (IRDye®800CW) preadsorbed (ab216772) and Goat Anti-Rabbit IgG H&L (IRDye®680RD) preadsorbed (ab216777) secondary antibodies at 1 in 20000 dilution for 1 hour at room temperature before imaging.



Allele-1: 1 bp insertion in exon 1.



Allele-2: 2 bp insertion in exon 1.



Cell Culture - Human FTL knockout HeLa cell line (ab265534)

Representative images of FTL knockout HeLa cells, low and high confluency examples (top left and right respectively) and wild-type HeLa cells, low and high confluency (bottom left and right respectively) showing typical adherent, epithelial-like morphology. Images were captured at 10X magnification using a EVOS XL Core microscope.

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