

Human TP53INP2 knockout HEK-293T cell line ab266474

1 Image

Overview

| | |
|-----------------------------|---|
| Product name | Human TP53INP2 knockout HEK-293T cell line |
| Parental Cell Line | HEK293T |
| Organism | Human |
| Mutation description | Knockout achieved by using CRISPR/Cas9, Homozygous: 1 bp insertion in exon 4 |
| Passage number | <20 |
| Knockout validation | Sanger Sequencing |
| Biosafety level | 2 |
| General notes | <p>Recommended control: Human wild-type HEK293T cell line (ab255449). Please note a wild-type cell line is not automatically included with a knockout cell line order, if required please add recommended wild-type cell line at no additional cost using the code WILDTYPE-TMTK1.</p> <p>Cryopreservation cell medium: Cell Freezing Medium-DMSO Serum free media, contains 8.7% DMSO in MEM supplemented with methyl cellulose.</p> <p>Culture medium: DMEM (High Glucose) + 10% FBS</p> <p>Initial handling guidelines: Upon arrival, the vial should be stored in liquid nitrogen vapor phase and not at -80°C. Storage at -80°C may result in loss of viability.</p> <ol style="list-style-type: none">1. Thaw the vial in 37°C water bath for approximately 1-2 minutes.2. Transfer the cell suspension (0.8 mL) to a 15 mL/50 mL conical sterile polypropylene centrifuge tube containing 8.4 mL pre-warmed culture medium, wash vial with an additional 0.8 mL culture medium (total volume 10 mL) to collect remaining cells, and centrifuge at 201 x g (rcf) for 5 minutes at room temperature. 10 mL represents minimum recommended dilution. 20 mL represents maximum recommended dilution.3. Resuspend the cell pellet in 5 mL pre-warmed culture medium and count using a haemocytometer or alternative cell counting method. Based on cell count, seed cells in an appropriate cell culture flask at a density of 2×10^4 cells/cm². Seeding density is given as a guide only and should be scaled to align with individual lab schedules.4. Incubate the culture at 37°C incubator with 5% CO₂. Cultures should be monitored daily. <p>Subculture guidelines:</p> <p>All seeding densities should be based on cell counts gained by established methods. A guide seeding density of 2×10^4 cells/cm² is recommended.</p> <p>A partial media change 24 hours prior to subculture may be helpful to encourage growth, if required.</p> <p>Cells should be passaged when they have achieved 80-90% confluence.</p> |

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We will provide viable cells that proliferate on revival.

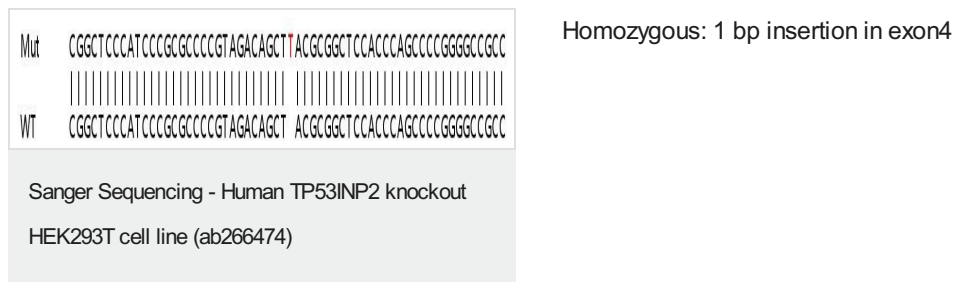
Properties

| | |
|-----------------------------|--|
| Number of cells | 1 x 10 ⁶ cells/vial, 1 mL |
| Adherent /Suspension | Adherent |
| Tissue | Kidney |
| Cell type | epithelial |
| STR Analysis | Amelogenin X D5S818: 8, 9 D13S317: 12, 14 D7S820: 11 D16S539: 9, 13 vWA: 16, 19 TH01: 7, 9.3 TPOX: 11 CSF1PO: 11, 12 |
| Mycoplasma free | Yes |
| Storage instructions | Shipped on Dry Ice. Store in liquid nitrogen. |
| Storage buffer | Constituents: 8.7% Dimethylsulfoxide, 2% Cellulose, methyl ether |

Target

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|------------------------------|----------|
| Cellular localization | Nucleus. |
|------------------------------|----------|

Images



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