abcam

Product datasheet

Human VIM (Vimentin) knockout HeLa cell line ab255446

3 Images

Overview

Product name Human VIM (Vimentin) knockout HeLa cell line

Parental Cell Line HeLa
Organism Human

Mutation description Knockout achieved by using CRISPR/Cas9, Homozygous: Insertion of the selection cassette in

exon 2

Passage number <20

Knockout validation Sanger Sequencing, Western Blot (WB)

Tested applications Suitable for: WB

Biosafety level 2

General notesRecommended control: Human wild-type HeLa cell line (<u>ab255448</u>). Please note a wild-type

cell line is not automatically included with a knockout cell line order, if required please add

cell line is not automatically included with a knockout cell line order, if required please add recommended wild-type cell line at no additional cost using the code WILDTYPE-TMTK1.

Cryopreservation cell medium: Cell Freezing Medium-DMSO Serum free media, contains

8.7% DMSO in MEM supplemented with methyl cellulose.

Culture medium: DMEM (High Glucose) + 10% FBS

Initial handling guidelines: Upon arrival, the vial should be stored in liquid nitrogen vapor phase and not at -80°C. Storage at -80°C may result in loss of viability.

- 1. Thaw the vial in 37°C water bath for approximately 1-2 minutes.
- 2. Transfer the cell suspension (0.8 mL) to a 15 mL/50 mL conical sterile polypropylene centrifuge tube containing 8.4 mL pre-warmed culture medium, wash vial with an additional 0.8 mL culture medium (total volume 10 mL) to collect remaining cells, and centrifuge at 201 x g (rcf) for 5 minutes at room temperature. 10 mL represents minimum recommended dilution. 20 mL represents maximum recommended dilution.
- 3. Resuspend the cell pellet in 5 mL pre-warmed culture medium and count using a haemocytometer or alternative cell counting method. Based on cell count, seed cells in an appropriate cell culture flask at a density of 2x10⁴ cells/cm². Seeding density is given as a guide only and should be scaled to align with individual lab schedules.
- 4. Incubate the culture at 37°C incubator with 5% CO₂. Cultures should be monitored daily.

Subculture guidelines:

All seeding densities should be based on cell counts gained by established methods.

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A guide seeding density of 2x10⁴ cells/cm² is recommended.

A partial media change 24 hours prior to subculture may be helpful to encourage growth, if required.

Cells should be passaged when they have achieved 80-90% confluence.

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We will provide viable cells that proliferate on revival.

Properties

1 x 10⁶ cells/vial, 1 mL Number of cells

Adherent/Suspension Adherent **Tissue** Cervix Cell type epithelial

Disease Adenocarcinoma

Gender Female

STR Analysis Amelogenin X D5S818: 11, 12 D13S317: 12, 13.3 D7S820: 8, 12 D16S539: 9, 10 vWA: 16, 18

TH01: 7 TPOX: 8, 12 CSF1PO: 9, 10

Mycoplasma free Yes

Storage instructions Shipped on Dry Ice. Store in liquid nitrogen.

Storage buffer Constituents: 8.7% Dimethylsulfoxide, 2% Cellulose, methyl ether

Target

Function Vimentins are class-Ill intermediate filaments found in various non-epithelial cells, especially

mesenchymal cells. Vimentin is attached to the nucleus, endoplasmic reticulum, and

mitochondria, either laterally or terminally.

Involved with LARP6 in the stabilization of type I collagen mRNAs for CO1A1 and CO1A2.

Tissue specificity Highly expressed in fibroblasts, some expression in T- and B-lymphocytes, and little or no

expression in Burkitt's lymphoma cell lines. Expressed in many hormone-independent mammary

carcinoma cell lines.

Involvement in disease Cataract 30

Sequence similarities Belongs to the intermediate filament family.

Domain The central alpha-helical coiled-coil rod region mediates elementary homodimerization.

The [IL]-x-C-x-x-[DE] motif is a proposed target motif for cysteine S-nitrosylation mediated by the

iNOS-S100A8/A9 transnitrosylase complex.

Post-translational

Filament disassembly during mitosis is promoted by phosphorylation at Ser-55 as well as by modifications

nestin (By similarity). One of the most prominent phosphoproteins in various cells of mesenchymal origin. Phosphorylation is enhanced during cell division, at which time vimentin filaments are

significantly reorganized. Phosphorylation by PKN1 inhibits the formation of filaments.

Phosphorylated at Ser-56 by CDK5 during neutrophil secretion in the cytoplasm. Phosphorylated

by STK33.

O-glycosylated during cytokinesis at sites identical or close to phosphorylation sites, this

interferes with the phosphorylation status.

 $S-nitrosylation\ is\ induced\ by\ interferon-gamma\ and\ oxidatively-modified\ low-densitity\ lipoprotein$

(LDL(ox)) possibly implicating the iNOS-S100A8/9 transnitrosylase complex.

Cellular localization

Cytoplasm.

Form

Vimentin is found in connective tissue and in the cytoskeleton.

Applications

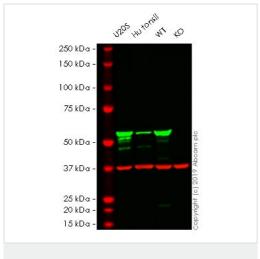
The Abpromise guarantee

Our <u>Abpromise guarantee</u> covers the use of ab255446 in the following tested applications.

The application notes include recommended starting dilutions; optimal dilutions/concentrations should be determined by the end user.

Application	Abreviews	Notes
WB		Use at an assay dependent concentration. Predicted molecular weight: 53 kDa.

Images



Western blot - Human VIM (Vimentin) knockout HeLa cell line (ab255446) All lanes: Anti-Vimentin antibody [V9] - Cytoskeleton Marker

(ab8069) at 1 µg/ml

Lane 1 : U-2 OS (Human bone osteosarcoma epithelial cell line)

cell lysate

Lane 2: Human tonsil cell lysate

Lane 3: Wild-type HeLa cell lysate

Lane 4: VIM knockout HeLa cell lysate

Lysates/proteins at 20 µg per lane.

Secondary

All lanes : Goat anti-Mouse IgG H&L (IRDye® 800CW)

preadsorbed (ab216772) at 1/20000 dilution

Performed under reducing conditions.

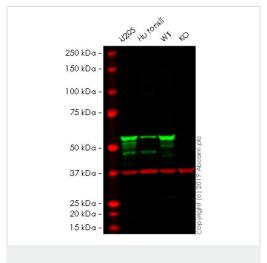
Predicted band size: 53 kDa

Additional bands at: 50 kDa (possible Loading Control)

Lanes 1 - 4: Merged signal (red and green). Green - <u>ab8069</u> observed at 53 kDa. Red - loading control, <u>ab181602</u> observed at 37 kDa.

ab8069 was shown to react with Vimentin in wild-type HeLa cells.

Loss of signal was observed when knockout cell line ab255446 (knockout cell lysate <u>ab263775</u>) was used. Wild-type and Vimentin knockout samples were subjected to SDS-PAGE. <u>ab8069</u> and Anti-GAPDH antibody [EPR16891] - Loading Control (<u>ab181602</u>) were incubated overnight at 4°C at 1 μ g/ml and 1 in 20000 dilution respectively. Blots were developed with Goat anti-Mouse lgG H&L (IRDye® 800CW) preadsorbed (<u>ab216772</u>) and Goat Anti-Rabbit lgG H&L (IRDye® 680RD) preadsorbed (<u>ab216777</u>) secondary antibodies at 1 in 20000 dilution for 1 hour at room temperature before imaging.



Western blot - Human VIM (Vimentin) knockout HeLa cell line (ab255446) **All lanes :** Anti-Vimentin antibody [SP20] (ab16700) at 1/100 dilution

Lane 1: U20S cell lysate

Lane 2: Human tonsil cell lysate

Lane 3: Wild-type HeLa cell lysate

Lane 4: VIM knockout HeLa cell lysate

Lysates/proteins at 20 µg per lane.

Secondary

All lanes : Goat anti-Rabbit lgG H&L (IRDye® 800CW) preadsorbed (<u>ab216773</u>) at 1/20000 dilution

Performed under reducing conditions.

Predicted band size: 53 kDa

Additional bands at: 37 kDa (possible Loading Control)

Lanes 1 - 4: Merged signal (red and green). Green - <u>ab16700</u> observed at 53 kDa. Red - loading control, <u>ab8245</u> observed at 37 kDa.

<u>ab16700</u> was shown to react with Vimentin in wild-type HeLa cells. Loss of signal was observed when knockout cell line ab255446 (knockout cell lysate <u>ab263775</u>) was used. Wild-type and Vimentin knockout samples were subjected to SDS-PAGE. <u>ab16700</u> and Anti-GAPDH antibody [6C5] - Loading Control (<u>ab8245</u>) were incubated overnight at 4°C at 1 in 100 dilution and 1 in 20000

dilution respectively. Blots were developed with Goat anti-Rabbit lgG H&L (IRDye[®] 800CW) preadsorbed (<u>ab216773</u>) and Goat anti-Mouse lgG H&L (IRDye[®] 680RD) preadsorbed (<u>ab216776</u>) secondary antibodies at 1 in 20000 dilution for 1 hour at room temperature before imaging.



Homozygous: Insertion of the selection cassette in exon 2.

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