ab139460

PDE Activity Assay Kit (Colorimetric)

Instructions for Use

For the screening of inhibitors and modulators of cyclic nucleotide phosphodiesterase activity.

View kit datasheet: www.abcam.com/ab139460
(use www.abcam.cn/ab139460 for China, or www.abcam.co.jp/ab139460 for Japan)

This product is for research use only and is not intended for diagnostic use.
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1. Principle of the Assay

Abcam PDE Activity Assay Kit (Colorimetric) (ab139460) is a colorimetric, non-radioactive assay designed in a microplate format. It may be used to screen inhibitors and modulators of cyclic nucleotide phosphodiesterase activity.

The basis for the assay is the cleavage of cAMP or cGMP by a cyclic nucleotide phosphodiesterase. The 5′-nucleotide released is further cleaved into the nucleoside and phosphate by the enzyme 5′-nucleotidase. The phosphate released due to enzymatic cleavage is quantified using Green Assay Reagent in a modified Malachite Green assay. The kit includes Type I cyclic AMP phosphodiesterase (PDE) for validation purposes.

A nonspecific cyclic nucleotide phosphodiesterase inhibitor, 3-isobutyl-1-methylxanthine (IBMX) is included as a test control for inhibitor screening. It has an IC$_{50}$ of approximately 25 μM.

It is important to ensure that compounds active in the cyclic nucleotide phosphodiesterase assay do not inhibit the activity of the 5′-nucleotidase. This can be ascertained by using 5′-AMP rather than 3′,5′-cAMP as the substrate. For further information, see section 7.B. “Preparing a Standard Curve”.
This kit is sold based on number of tests. A ‘test’ simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions.

The assay offers the following advantages:

1) Non-radioactive
2) Convenient one step detection
3) Microplate format
2. Protocol Summary

Desalt samples

Prepare standard curve

Dilute standards

Prepare standard wells

Add 5’-nucleotidase to each well and mix thoroughly

Incubate for 30 minutes at 30°C

Terminate reaction by adding Green Assay Reagent
**Prepare time course/linearity assay**

Dilute cAMP substrate

Add cAMP substrate to appropriate wells

Add assay buffer to each well

Add 5’-nucleotidase to each well and mix thoroughly

Dilute PDE enzyme

Add PDE enzyme to each well at 5 minute intervals

Terminate reaction by adding Green Assay Reagent

**Prepare test sample/inhibitor assay**

Prepare samples containing PDE enzyme, substrate and test compound. Include IBMX (optional)

Incubate samples at appropriate temperature and time

Terminate reaction by adding Green Assay Reagent
## 3. Materials Supplied

<table>
<thead>
<tr>
<th>Item</th>
<th>Quantity</th>
<th>Storage</th>
</tr>
</thead>
<tbody>
<tr>
<td>PDE Enzyme (from bovine brain) (lyophilized) (4 U per vial)</td>
<td>5 vials</td>
<td>-80°C</td>
</tr>
<tr>
<td>5’-Nucleotidase (5 kU/µL) (from <em>Crotalus atrox</em> venom)</td>
<td>1 x 1 mL</td>
<td>-80°C</td>
</tr>
<tr>
<td>3’,5’-cAMP Substrate (1 mM)</td>
<td>1 x 2 mL</td>
<td>-80°C</td>
</tr>
<tr>
<td>3’,5’-cGMP Substrate (1 mM)</td>
<td>1 x 2 mL</td>
<td>-80°C</td>
</tr>
<tr>
<td>PDE Assay Buffer</td>
<td>1 x 40 mL</td>
<td>-80°C</td>
</tr>
<tr>
<td>Green Assay Reagent</td>
<td>1 x 20 mL</td>
<td>4°C</td>
</tr>
<tr>
<td>5’-AMP Standard (100 µM)</td>
<td>1 x 1 mL</td>
<td>-80°C</td>
</tr>
<tr>
<td>5’-GMP Standard (100 µM)</td>
<td>1 x 1 mL</td>
<td>-80°C</td>
</tr>
<tr>
<td>PDE Inhibitor (IBMX) (200 µM)</td>
<td>1 x 200 µL</td>
<td>-80°C</td>
</tr>
<tr>
<td>Desalting Column</td>
<td>1</td>
<td>4°C</td>
</tr>
<tr>
<td>Desalting Resin</td>
<td>1 x 1g</td>
<td>4°C</td>
</tr>
<tr>
<td>96-well Clear Microplate (½ Volume)</td>
<td>1</td>
<td>4°C</td>
</tr>
</tbody>
</table>
4. Storage and Stability

- Please note that all components, with the exception of the Green Assay Reagent, Desalting Column, Desalting Resin, PDE Enzyme, and the 5’-nucleotidase can be stored at either -80°C or -20°C.
- The PDE Enzyme and 5’-Nucleotidase must be stored at -80°C.
- Because its stability when frozen in solution is poor, the PDE enzyme is provided as 5 lyophilized aliquots. For highest activity, we recommend that each lyophilized aliquot be dissolved and used for one day’s experiments (See Section 7.A. Reagent Preparation).

- The 5’-nucleotidase is used undiluted at 10 μL per well. It is stable for up to 6 freeze/thaw cycles (snap freezing with liquid N₂ or dry ice/ethanol), but it may be desirable to make several aliquots if more numerous freeze/thaws are planned.

- One U of 5’-nucleotidase will release 1 pmol phosphate per minute from 200 μM 5’-AMP, at 30°C in a reaction buffer of 10 mM Tris-HCl, pH 7.4, 0.2 mM MgCl₂.

- One U of PDE enzyme = 1 nmol 3’ 5’-cAMP to 5’-AMP per minute under the conditions of the linearity assay
The Green Assay Reagent is a highly sensitive phosphate detection solution. Free phosphate present on labware and in reagent solutions will greatly increase the background absorbance of the assay. This is detected visually as a change in color from yellow to green. Detergents used to clean labware may contain high levels of phosphate. Use caution by either rinsing labware with dH$_2$O or employ unused plasticware.

5. Materials Required, Not Supplied

- Microplate reader capable of measuring $A_{620}$ to $\geq$3-decimal accuracy.

- Pipettes capable of pipetting 10-100 µL accurately.

- Multi-channel pipetman capable of pipetting 100 µl (optional).

- Ice bucket to keep reagents cold until use.
6. Sample Preparation

Note: The following procedures are intended only as a guideline. The optimal experimental conditions will vary depending on the parameters being investigated, and must be determined by the individual user.

Desalting tissue samples by gel filtration

Note: This procedure is intended to remove excess phosphate and nucleotides (which are slowly hydrolyzed to release free phosphate in the presence of the Green Assay Reagent) in the high speed supernatant (HSS) extract.

1. Rehydrate Desalting Resin in a 50 mL conical tube by adding 20 mL of phosphate free dH₂O and vortexing briefly. Allow to set for 4 hours at RT or overnight at 4°C.
2. Decant the dH₂O carefully, then add fresh dH₂O at a 1:1 ratio to the rehydrated resin (~10 mL).
3. Add rehydrated resin to the Desalting Column to obtain a 5 mL settled-bed volume (~5.5 cm bed height). Remove tip from column and allow dH₂O to drain by gravity.
4. Equilibrate column by adding 8 mL of assay buffer and allow to drain by gravity.
5. Place column in a 15 mL centrifuge tube. Centrifuge at 800 x g for 3 min at 4°C to displace column buffer. Discard flow-through buffer.

6. Place column in a clean 15 mL centrifuge tube.

7. Add up to 350 μL sample to column.

8. Centrifuge at 800 x g for 3 min. Save extract flow-through. This is the desalted cell lysate material to be tested for PDE activity below.

9. Freeze sample immediately at -80°C.

Note: The effective removal of phosphate/nucleotides from the extract should be tested qualitatively by adding 100 μL Green Assay Reagent to 1 μL extract, and a separate sample of 1 μL dH2O. If no phosphate/nucleotides are present, both samples should remain yellow in color over a time period of 30 min @ RT. The development of a visible green color indicates phosphate contamination, which must be eliminated from the samples before proceeding further.

The rehydrated resin can be stored at 4°C until use. Please note that the volume of Desalting Resin provided in this kit allows user to fill the column twice (5mL settled-bed volume each). In theory, the rehydrated resin can be reused for multiple times (or samples) *. The column will lose efficiency in removing free phosphate gradually after each use. The number of samples that can be desalted by the resin will vary depending on the sample types and amount of phosphate.
present in the samples. In general, an average of 5-7 samples can be processed with each 5mL settled-bed volume of rehydrated resin, ie an average of 10-14 samples can be desalted per kit.

*When reusing the rehydrated resin, the column should be rinsed with 4-5 volumes of phosphate-free dH2O, and then equilibrated with 8 mL of a solution of 10mM Tris-hydrochloric acid, pH 7.4 before loading the next sample.
7. Assay Protocol

A. Reagent Preparation
1. Thaw assay buffer, 5’-nucleotidase, the 3’,5’-cAMP substrate, IBMX inhibitor, 5’-AMP and/or 5’-GMP standard. Store all on ice.
2. Prepare 20 U/mL solution of PDE by adding 200 µL of cold assay buffer to one of the vials of lyophilized enzyme. Store on ice. Note that each of the lyophilized PDE aliquots provided are intended for use in one day’s assays only. The enzyme solution may lose substantial activity from freezing/thawing and frozen storage.
3. Warm Green Assay Reagent to room temperature.

B. Preparing a Standard Curve
1. Prepare two dilutions in PDE Assay buffer, 75 µM and 50 µM, using either the 5’-AMP or 5’-GMP Standard. For example, bring aliquots of 150 µL and 100 µL 5’-AMP Standard (100 µM) and to 200 µL with assay buffer.
2. Using PDE Assay Buffer, prepare two sets of 1:1 serial dilutions of the 75 µM and 50 µM 5’-AMP standard, plus an assay buffer blank (40 µL per well). Concentrations of 75, 50, 37.5, 25, 18.75, 12.5, 6.25 µM correspond to 3, 2, 1.5, 1.0, 0.75, 0.50 and 0.25 nmol 5’-AMP or 5’-GMP (see Table 1):
a) Add 80 µL of 75 µM 5’-AMP or 5’-GMP standard (Step 1) to well A of assay plate and 80 µL of 50 µM standard to well B.

b) Add 40 µL 1X assay buffer to wells C through H.

c) Remove 40 µL from well A and add it to well C. Mix thoroughly by pipetting up and down several times.

d) Remove 40 µL from well C and add it to well E. Mix well E thoroughly and then remove 40 µL and discard.

e) Remove 40 µL from well B and add it to well D. Mix thoroughly by pipetting up and down several times. Repeat this process moving and mixing 40 µL from well D to F and then from F to G. Remove and discard 40 µL from well G. DO NOT PROCEED TO THE BLANK WELL ‘H’.

f) Add 10 µL of 5’-nucleotidase (undiluted, 5 kU/µL) to each well and mix thoroughly.

g) Incubate at 30°C for 30 minutes.

h) Proceed to “Terminating Reactions” (Section 7.E).
Table 1. Example of standard curve and time course/linearity microplate samples.

<table>
<thead>
<tr>
<th>Sample Well</th>
<th>5'-AMP/5'-GMP Standard Curve nmol 5'-AMP or 5'-GMP (Columns 1,2)</th>
<th>Time course Min. (Columns 3,4)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>3.0</td>
<td>90</td>
</tr>
<tr>
<td>B</td>
<td>2.0</td>
<td>60</td>
</tr>
<tr>
<td>C</td>
<td>1.5</td>
<td>45</td>
</tr>
<tr>
<td>D</td>
<td>1.0</td>
<td>30</td>
</tr>
<tr>
<td>E</td>
<td>0.75</td>
<td>20</td>
</tr>
<tr>
<td>F</td>
<td>0.50</td>
<td>10</td>
</tr>
<tr>
<td>G</td>
<td>0.25</td>
<td>5</td>
</tr>
<tr>
<td>H</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

For highest accuracy, perform all samples in duplicate. See Figures 1 and 2 for example results.
C. Preparing a Time Course/Linearity Assay

1. Dilute cAMP substrate to 0.5 mM with assay buffer.
2. Add 20 µL of cAMP substrate (0.5 mM) to appropriate wells. The final substrate concentration will be 200 µM.
3. Add 15 µL of assay buffer to each well.
4. Add 10 µL of 5’-nucleotidase (undiluted, 5 kU/µL) per well.
5. Designate a reaction time to each well (e.g.: 90, 60, 45, 30, 20, 10, 5 and 0 min). See Table 1.
6. Equilibrate microplate to reaction temperature (e.g.: 30°C).
7. Prepare PDE enzyme at 20 U/mL (See Section 7.A. Reagent Preparation). Dilute with assay buffer to 4 U/mL, making enough for the assays planned. Each well will receive 5 µL. Store dilution on ice.
8. Start reactions by addition of 5 µL of PDE enzyme. Total PDE enzyme= 20 mU/well. Make the additions in the reverse time order such that all incubations end at the same time (e.g.: Add 90 min time pt. at t=0; add 5 min at t=85 min, etc.). The total reaction volume=50 µL.
D. Preparing a Test Sample/Inhibitor Assay

1. Prepare samples containing PDE enzyme, substrate and test compound dissolved in PDE Assay Buffer as listed in Table 2. Include the PDE Inhibitor if desired.

2. Incubate samples at appropriate temperature (e.g. 37°C) and time (e.g.: 60 min).

Table 2: Example of Test Samples/Inhibitor Assay Microplate Samples

<table>
<thead>
<tr>
<th>Substrate (0.5 mM)</th>
<th>PDE Assay Buffer</th>
<th>5'-Nase (5 kU/µL)</th>
<th>Test compound</th>
<th>PDE Enzyme (4mU/ µL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>20 µL</td>
<td>15 µL</td>
<td>10 µL</td>
<td>0 µL</td>
</tr>
<tr>
<td>Test</td>
<td>20 µL</td>
<td>5 µL</td>
<td>10 µL</td>
<td>10 µL</td>
</tr>
<tr>
<td>PDE Inhibitor</td>
<td>20 µL</td>
<td>5 µL</td>
<td>10 µL</td>
<td>10 µL</td>
</tr>
</tbody>
</table>

3. To confirm that an apparent PDE inhibitor does not interfere with the release of phosphate by 5'-nucleotidase, test additional wells using 50 µM 5'-AMP or 5'-GMP standard and the inhibitor(s) in question. Compare the results with the 50 µM standard curve well.
E. Terminating Reactions

1. After incubating wells for desired duration, terminate reactions by addition of 100 µL Green Assay Reagent. Agitate plate or triturate wells gently to mix. Note: Avoid production of air bubbles in the wells.

2. Allow color to develop for 20-30 minutes. Be careful to assure samples spend approximately the same time with the reagent before reading on the microplate reader.

3. Read OD$_{620nm}$ on microtiter-plate reader. Note: Retain microtiter plate for future use of unused wells.
8. Data Analysis

A. 5’-AMP or 5’-GMP Standard Curve

1. Plot standard curve data as OD_{620nm} versus nmol 5’-AMP or 5’-GMP (see Fig. 1).
2. Fit a line to the plotted data using an appropriate linear regression program.
3. Rearrange the equation for best-fit line to solve for nmol of 5’-AMP or 5’-GMP in terms of OD_{620nm}.

\[
5’-\text{AMP released} = \frac{(\text{OD}_{620\text{nm}} - \text{y-intercept})}{\text{slope}}
\]

**SAMPLE CALCULATION:**

Best-fit eqn.: \( \text{OD}_{620\text{nm}} = 0.232(\text{nmol 5’-AMP}) + 0.0709 \)

Rearranged eqn.: \( \text{nmol 5’-AMP} = \frac{(\text{OD}_{620\text{nm}} - 0.0709)}{0.232} \)

Example: An unknown produces an \( \text{OD}_{620\text{nm}} = 0.400 \)

\[
5’-\text{AMP released} = \frac{(0.400 - 0.0709)}{0.232} = 1.42 \text{ nmol}
\]

4. Substitute \( \text{OD}_{620\text{nm}} \) data obtained from experimental samples (e.g. a PDE reaction) into the rearranged equation to obtain the nmol of 5’-AMP or 5’GMP produced.
Figure 1. Standard Curve for 5'AMP. Duplicate wells of 5'-AMP dilutions were prepared as described (see Section 8.B). Phosphate was released from 5'-AMP by incubation with 5'-nucleotidase (50 kU/well, 30°C, 30 min.) and the reaction terminated by addition of Green Assay Reagent (100 µL/well). After 30 min., the phosphate-dependent color reaction was measured by reading OD$_{620\text{nm}}$ in a microplate-reading spectrophotometer.

B. Determining the linear range of a PDE reaction time course

Assays for inhibitor screening are most sensitive when the results reflect the initial rate of the enzyme. It is therefore important to choose an incubation time that lies within the initial, linear part of the reaction progress curve. The time courses depicted in Figure 2, remain linear over the course of 45 min. Reasons for the departure from linearity late in a time course may include depletion of substrate, product inhibition and instability of the enzyme. It should also be noted that standard curves for cAMP or cGMP can
themselves become non-linear above 3 nmol (OD_{620nm} > 0.8). PDE incubation times that generate more than 3 nmol of cAMP or cGMP should therefore be avoided.

Figure 2. Time Course of cAMP Hydrolysis by PDE, Inhibition by IBMX. PDE enzyme (20 mU/well) was incubated with cAMP (200 μM) and 5'-nucleotidase (50 kU/well) with or without the inhibitor IBMX (40 μM) at 30°C for the indicated times. Reactions were terminated by addition of 100 μL of Green Assay Reagent and OD_{620nm} read 30 min. later. A cAMP standard curve (see Fig. 1) may be used to convert OD_{620nm} data to nmol of 5’-AMP.
## 9. Troubleshooting

<table>
<thead>
<tr>
<th>Problem</th>
<th>Reason</th>
<th>Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td>Assay not working</td>
<td>Assay buffer at wrong temperature</td>
<td>Assay buffer must not be chilled – needs to be at RT</td>
</tr>
<tr>
<td>Protocol step missed</td>
<td></td>
<td>Re-read and follow the protocol exactly</td>
</tr>
<tr>
<td>Plate read at incorrect wavelength</td>
<td></td>
<td>Ensure you are using appropriate reader and filter settings</td>
</tr>
<tr>
<td>Unsuitable microtiter plate for assay</td>
<td></td>
<td>Fluorescence: Black plates (clear bottoms); Luminescence: White plates; Colorimetry: Clear plates. If critical, protocol will indicate whether to use flat- or U-shaped wells</td>
</tr>
<tr>
<td>Problem</td>
<td>Reason</td>
<td>Solution</td>
</tr>
<tr>
<td>------------------------------------------</td>
<td>-------------------------------------</td>
<td>--------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Unexpected results</td>
<td>Measured at wrong wavelength</td>
<td>Ensure you are using appropriate reader and filter settings</td>
</tr>
<tr>
<td></td>
<td>Unsuitable sample type</td>
<td>Refer to datasheet for details about incompatible samples</td>
</tr>
<tr>
<td></td>
<td>Sample readings are outside linear range</td>
<td>Concentrate/ dilute samples to be in linear range</td>
</tr>
<tr>
<td>Samples with inconsistent readings</td>
<td>Unsuitable sample type</td>
<td>Refer to datasheet for details about incompatible samples</td>
</tr>
<tr>
<td></td>
<td>Too many freeze/thaw cycles</td>
<td>Aliquot samples to reduce the number of freeze/thaw cycles</td>
</tr>
<tr>
<td>Problem</td>
<td>Reason</td>
<td>Solution</td>
</tr>
<tr>
<td>-------------------------------------------</td>
<td>-------------------------------</td>
<td>---------------------------------------------------</td>
</tr>
<tr>
<td>Samples are too old or incorrectly stored</td>
<td>Use fresh made samples and store at recommended temperature until use</td>
<td></td>
</tr>
<tr>
<td>Not fully thawed kit components</td>
<td>Wait for components to thaw completely and gently mix prior use</td>
<td></td>
</tr>
<tr>
<td>Reagents sitting for extended periods on ice</td>
<td>Try to prepare a fresh reaction mix prior to each use</td>
<td></td>
</tr>
<tr>
<td>Incorrect amounts used</td>
<td>Check pipette is calibrated correctly (always use smallest volume pipette that can pipette entire volume)</td>
<td></td>
</tr>
<tr>
<td>Problem</td>
<td>Reason</td>
<td>Solution</td>
</tr>
<tr>
<td>---------------------------------</td>
<td>---------------------------------------</td>
<td>---------------------------------------------------</td>
</tr>
<tr>
<td>Standard curve is not linear</td>
<td>Not fully thawed kit components</td>
<td>Wait for components to thaw completely and gently mix prior use</td>
</tr>
<tr>
<td>Pipetting errors when setting up the standard curve</td>
<td>Try not to pipette too small volumes</td>
<td></td>
</tr>
<tr>
<td>Incorrect pipetting when setting up the reaction mix</td>
<td>Always prepare a master mix</td>
<td></td>
</tr>
<tr>
<td>Air bubbles in wells</td>
<td>Air bubbles will interfere with readings; try to avoid producing air bubbles and always remove bubbles prior to reading plates</td>
<td></td>
</tr>
</tbody>
</table>
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