ab139488 –
CytoPainter
Lysosome/ER/Nuclear
Staining Reagent
(Fluorometric)

Instructions for Use

Designed to detect lysosomes, endoplasmic reticulum and nucleic acids by microscopy.

This product is for research use only and is not intended for diagnostic use.
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1. Introduction

ab139488 CytoPainter Lysosome/ER/Nuclear Staining Reagent is optimal for use in demanding cell analysis applications involving confocal microscopy, flow cytometry, microplate readers and HCS/HTS, where consistency and reproducibility are required.

2. Product Overview

ab139488 contains a mixture of cell-permeable red fluorescent lysosomal dye, green fluorescent endoplasmic reticulum dye and blue fluorescent nucleic acid dye. The staining pattern arising from the combination of these three dyes permits visualization of the target organelles by fluorescence/confocal microscopy. The reagent, supplied as a 500X solution, is sufficient for 1000 microscopy assays. The single-tube format makes this multi-organelle stain reagent easy to use.
3. Components and Storage

A. Kit Contents

<table>
<thead>
<tr>
<th>Item</th>
<th>Quantity</th>
<th>Storage Temperature</th>
</tr>
</thead>
<tbody>
<tr>
<td>Organelle Reagent II</td>
<td>200 µL</td>
<td>-80°C</td>
</tr>
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</table>

Reagents provided in the kit are sufficient for approximately 1000 microscopy assays using either live, adherent cells or cells in suspension.

B. Storage and Handling

Upon receipt, the kit should be stored -80°C, protected from light. Avoid repeated freezing and thawing.

C. Additional Materials Required

- Standard fluorescence microscope
- Calibrated, adjustable precision pipets, preferably with disposable plastic tips
- Adjustable speed centrifuge with swinging buckets (for suspension cultures)
- Glass microscope slides
- Glass cover slips (18 x 18 mm)
- Deionized water
- Anhydrous DMSO (optional).
- Growth medium (e.g. Dulbecco’s Modified Eagle medium, D-MEM)
- Paraformaldehyde (optional, for fixation)
- Triton X-100 (optional, for permeabilization)

4. Pre-Assay Preparation

NOTE: Allow all reagents to thaw at room temperature before starting with the procedures. Upon thawing, gently hand-mix or vortex the reagents prior to use to ensure a homogenous solution. Briefly centrifuge the vials at the time of first use, as well as for all subsequent uses, to gather the contents at the bottom of the tube.

A. Reagent Preparation

Mix 2 μL of Organelle Reagent II in 1 mL of buffer of choice. This volume is sufficient for 10 assays and may be scaled according to need.
5. Assay Protocol

Wide Field Fluorescence/Confocal Microscopy

A. Staining Live, Adherent Cells
1. Grow cells directly onto glass slides or polystyrene tissue culture plates until ~80% confluent via standard tissue culture practices.
2. Remove growth media.
3. Dispense the freshly diluted staining solution in a volume sufficient for covering the cell monolayer.
4. Protect samples from light and incubate for 30 minutes at 37°C.
5. Remove the excess staining solution and, if necessary, add a few drops of buffer to prevent the cells from drying out.
6. Cover cells with a glass cover slip and observe under a fluorescence/confocal microscope with a filter set for DAPI (Ex/Em: 350/470nm), Texas Red (Ex/Em: 540/605 nm) and GFP/FITC (Ex/Em: 488/514 nm).

B. Staining Live Cells Grown in Suspension
1. Grow the cells via standard tissue culture practices.
2. Collect about 1 x 10^5 cells. Centrifuge at 500 x g for 5 minutes. Remove supernatant.
3. Re-suspend cells in a volume of the freshly diluted staining solution sufficient for covering the cell pellet.
4. Protect the samples from light and incubate for 30 minutes at 37°C.
5. Centrifuge at 500 x g for 5 minutes. Remove supernatant.
6. Re-suspend the cells in 100 µL buffer.
7. Plate 10-15 µL of cells on a glass slide.
8. Cover cells with a glass cover slip and observe under a fluorescence/confocal microscope with a filter set for DAPI (Ex/Em: 350/470nm), Texas Red (Ex/Em: 540/605 nm) and GFP/FITC (Ex/Em: 488/514 nm).

**Filter set selection**

The selection of optimal filter sets for a fluorescence microscopy application requires matching the optical filter specifications to the spectral characteristics of the dyes employed in the analysis. Consult the microscope or filter set manufacturer for assistance in selecting optimal filter sets for your microscope.

*Figure 1.* Live HeLa cells treated with Chloroquine and stained with Organelle Reagent II showing composite of (A) lysosomes in red, (B) endoplasmic reticula in green, and (C) nuclei in blue.
Wavelength Maxima:
Lysosomal (Red): Excitation: 568nm Emission: 667nm
Endoplasmic Reticulum (Green): Excitation: 440nm Emission: 565nm
Nuclear (Blue): Excitation: 350nm Emission: 461nm
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