Instructions for Use

For the quantitative measurement of Human MBL (Mannose-Binding Lectin) in serum, plasma and cell culture supernatants.

This product is for research use only and is not intended for diagnostic use.
Table of Contents

INTRODUCTION
1. BACKGROUND 2
2. ASSAY SUMMARY 3
3. PRECAUTIONS 4

GENERAL INFORMATION
4. STORAGE AND STABILITY 4
5. MATERIALS SUPPLIED 4
6. MATERIALS REQUIRED, NOT SUPPLIED 5
7. LIMITATIONS 5
8. TECHNICAL HINTS 5

ASSAY PREPARATION
9. REAGENT PREPARATION 7
10. STANDARD PREPARATIONS 8
11. SAMPLE PREPARATION 10
12. PLATE PREPARATION 10

ASSAY PROCEDURE
13. ASSAY PROCEDURE 11

DATA ANALYSIS
14. CALCULATIONS 12
15. TYPICAL DATA 13
16. TYPICAL SAMPLE VALUES 15
17. ASSAY SPECIFICITY 16

RESOURCES
18. TROUBLESHOOTING 17
19. NOTES 18
INTRODUCTION

1. BACKGROUND

Abcam’s MBL (Mannose-Binding Lectin) Human ELISA Kit (ab193709) is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of Human MBL in sera, plasma and cell culture supernatants.

This assay employs an antibody specific for Human MBL coated on a 96-well plate. Standards and samples are pipetted into the wells and the immobilized antibody captures MBL present in the samples. The wells are washed and biotinylated anti-Human MBL antibody is added. After washing away any unbound biotinylated antibody, an HRP-conjugated streptavidin is pipetted to the wells. After incubation, the wells are again washed, followed by the addition of a TMB substrate solution to the wells. Color will develop in proportion to the amount of MBL bound in each well. Addition of the Stop Solution will change the color from blue to yellow, and the intensity of the color is measured at 450 nm.
2. ASSAY SUMMARY

Prepare all reagents, samples and standards as instructed.

Add standard or sample to each well used. Incubate at room temperature.

Add prepared biotinylated antibody to each well. Incubate at room temperature.

Add prepared Streptavidin solution. Incubate at room temperature.

Add TMB One-Step Development Solution to each well. Incubate at room temperature. Add Stop Solution to each well. Read immediately.
3. PRECAUTIONS
Please read these instructions carefully prior to beginning the assay.
Modifications to the kit components or procedures may result in loss of performance.

4. STORAGE AND STABILITY
Store kit at -20°C immediately upon receipt.
Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in sections 9 & 10.

5. MATERIALS SUPPLIED

<table>
<thead>
<tr>
<th>Item</th>
<th>Amount</th>
<th>Storage Condition (Before Preparation)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pre-coated MBL Microplate (12 x 8 well strips)</td>
<td>96 wells</td>
<td>-20°C</td>
</tr>
<tr>
<td>20X Wash Buffer Concentrate</td>
<td>25 mL</td>
<td>-20°C</td>
</tr>
<tr>
<td>Human MBL Standard (Lyophilized)</td>
<td>2 vials</td>
<td>-20°C</td>
</tr>
<tr>
<td>Assay Diluent A</td>
<td>30 mL</td>
<td>-20°C</td>
</tr>
<tr>
<td>5X Assay Diluent B</td>
<td>15 mL</td>
<td>-20°C</td>
</tr>
<tr>
<td>Biotinylated Human MBL Detection Antibody (Lyophilized)</td>
<td>2 vials</td>
<td>-20°C</td>
</tr>
<tr>
<td>800X HRP-Streptavidin Concentrate</td>
<td>200 µL</td>
<td>-20°C</td>
</tr>
<tr>
<td>TMB One-Step Substrate Reagent</td>
<td>12 mL</td>
<td>-20°C</td>
</tr>
<tr>
<td>Stop Solution</td>
<td>8 mL</td>
<td>-20°C</td>
</tr>
</tbody>
</table>
6. **MATERIALS REQUIRED, NOT SUPPLIED**

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Microplate reader capable of measuring absorbance at 450 nm.
- Precision pipettes to deliver 2 μL to 1 mL volumes.
- Adjustable 1-25 mL pipettes for reagent preparation.
- 100 mL and 1 liter graduated cylinders.
- Absorbent paper.
- Distilled or deionized water.
- Log-log graph paper or computer and software for ELISA data analysis.
- Tubes to prepare standard or sample dilutions.

7. **LIMITATIONS**

- Do not mix or substitute reagents or materials from other kit lots or vendors.

8. **TECHNICAL HINTS**

- Samples which generate values that are greater than the most concentrated standard should be further diluted in the appropriate sample dilution buffer.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Completely aspirate all solutions and buffers during wash steps. When preparing your standards, it is critical to briefly centrifuge the vial first. The powder may adhere to the cap and not be included in
the standard solution resulting in an incorrect concentration. Be sure to dissolve the powder thoroughly when reconstituting. After adding Assay Diluent to the vial, we recommend inverting the tube a few times, then flick the tube a few times, and centrifuge briefly; repeat this procedure 3-4 times. This is an effective technique for thorough mixing of the standard without using excessive mechanical force.

- Do not vortex the standard during reconstitution, as this will destabilize the protein.
- Once your standard has been reconstituted, it should be used right away or else frozen for later use.
- Keep the standard dilutions on ice during preparation, but the ELISA procedure should be done at room temperature.
- Be sure to discard the working standard dilutions after use – they do not store well.
- This kit is sold based on number of tests. A ‘test’ simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Scientific Support staff with any questions.
9. REAGENT PREPARATION

Equilibrate all reagents and samples to room temperature (18-25°C) prior to use.

9.1 **1X Assay Diluent B**
Dilute 5X Assay Diluent B 5-fold with deionized or distilled water before use.

9.2 **1X Wash Buffer**
If the 20X Wash Concentrate contains visible crystals, equilibrate to room temperature and mix gently until dissolved. Dilute 20 mL of 20X Wash Buffer Concentrate into 380 mL deionized or distilled water to yield 400 mL of 1X Wash Buffer.

9.3 **Detection Antibody MBL (biotinylated anti-Human MBL)**
Briefly centrifuge the Detection Antibody vial before use. Add 100 μL of 1X Assay Diluent B into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4°C for 5 days). The detection antibody concentrate should be diluted 80-fold with 1X Assay Diluent B and used in Assay Procedure.

9.4 **Assay Diluent A**
Ready to use

9.5 **1X HRP-Streptavidin Solution**
Briefly centrifuge the 800X HRP-Streptavidin concentrate vial and pipette up and down to mix gently before use. The 800X HRP-Streptavidin concentrate should be diluted 800-fold with 1X Assay Diluent B.

For example: Briefly centrifuge the vial and pipette up and down to mix gently. Add 15 μL of HRP-Streptavidin concentrate into a tube with 12 mL 1X Assay Diluent B to prepare a 1X HRP-Streptavidin solution (do not store the diluted solution for next day use). Mix well.
10. STANDARD PREPARATIONS

- Prepare serially diluted standards immediately prior to use. Always prepare a fresh set of standards for every use.
- Standard (recombinant protein) should be stored at -20°C or -80°C (recommended at -80°C) after reconstitution.

10.1 Briefly centrifuge the vial of Human MBL Standard and then add 800 μL Assay Diluent A (for serum/plasma samples) or 1X Assay Diluent B (for cell culture supernatants) into the Human MBL Standard vial to prepare a 25 ng/mL Standard #1. Mix thoroughly but gently.

10.2 Label tubes #2-8.

10.3 Add 400 μL Assay Diluent A or 1X Assay Diluent B into tubes 2-8.

10.4 Prepare Standard #2 by adding 200 μL Standard #1 to tube #2. Mix thoroughly but gently.

10.5 Prepare Standard #3 by adding 200 μL from Standard #2 to tube #3. Mix thoroughly but gently.

10.6 Using the table below as a guide, prepare further serial dilutions.

10.7 Standard #8 contains no protein and is the Blank control.
Standard Dilution Preparation Table

<table>
<thead>
<tr>
<th>Standard #</th>
<th>Sample to Dilute</th>
<th>Volume to Dilute (µL)</th>
<th>Volume of Diluent (µL)</th>
<th>Starting Conc. (ng/mL)</th>
<th>Final Conc. (ng/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>See Step 10.1</td>
<td></td>
<td></td>
<td></td>
<td>25</td>
</tr>
<tr>
<td>2</td>
<td>Standard #1</td>
<td>200</td>
<td>400</td>
<td>25</td>
<td>8.333</td>
</tr>
<tr>
<td>3</td>
<td>Standard #2</td>
<td>200</td>
<td>400</td>
<td>8.333</td>
<td>2.778</td>
</tr>
<tr>
<td>4</td>
<td>Standard #3</td>
<td>200</td>
<td>400</td>
<td>2.778</td>
<td>0.926</td>
</tr>
<tr>
<td>5</td>
<td>Standard #4</td>
<td>200</td>
<td>400</td>
<td>0.926</td>
<td>0.309</td>
</tr>
<tr>
<td>6</td>
<td>Standard #5</td>
<td>200</td>
<td>400</td>
<td>0.309</td>
<td>0.103</td>
</tr>
<tr>
<td>7</td>
<td>Standard #6</td>
<td>200</td>
<td>400</td>
<td>0.103</td>
<td>0.034</td>
</tr>
<tr>
<td>8 (Blank)</td>
<td>none</td>
<td>-</td>
<td>400</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>
11. SAMPLE PREPARATION

- If your samples need to be diluted, 1X Assay Diluent A should be used for dilution of sera/plasma samples. 1X Assay Diluent B should be used for dilution of culture supernatants and urine.
- Suggested dilution for normal serum/plasma: 800 fold.
- Please note that levels of the target protein may vary between different specimens. Optimal dilution factors for each sample must be determined by the investigator.

12. PLATE PREPARATION

- The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents.
- Unused well strips should be returned to the plate packet and stored at 4°C.
- For each assay performed, a minimum of 2 wells must be used as blanks, omitting primary antibody from well additions.
- For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates).
- Well effects have not been observed with this assay.
13. ASSAY PROCEDURE

- Equilibrate all materials and prepared reagents to room temperature (18 - 25°C) prior to use.

  It is recommended to assay all standards, controls and samples in duplicate.

13.1. Add 100 μL of each standard (see Standard Preparations, section) and sample into appropriate wells. Cover plate and incubate for 2.5 hours at room temperature or overnight at 4°C with gentle shaking.

13.2. Discard the solution and wash 4 times with 1X Wash Buffer. Wash by filling each well with 300 μL 1X Wash Buffer using a multi-channel pipette or automatic microplate washer. Complete removal of liquid at each step is essential for good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it by tapping it gently against clean paper towels.

13.3. Add 100 μL of the prepared biotinylated Human MBL Detection Antibody (see Reagent Preparation section) to each well. Incubate for 1 hour at room temperature with gentle shaking.

13.4. Discard the solution. Repeat the wash as in step 13.2.

13.5. Add 100 μL of prepared 1X HRP-Streptavidin solution (see Reagent Preparation section) to each well. Incubate for 45 minutes at room temperature with gentle shaking.

13.6. Discard the solution. Repeat the wash as in step 13.2.

13.7. Add 100 μL of TMB One-Step Substrate Reagent to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.

13.8. Add 50 μL of Stop Solution to each well. Read at 450 nm immediately.
14. **CALCULATIONS**

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.
15. **TYPICAL DATA**

**TYPICAL STANDARD CURVE** – Data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.

![Graph](image)

**Figure 1.** Example of typical Human MBL standard curve in Assay Diluent A (used for serum/plasma standards). The standard curve was prepared as described in Section 10.
Figure 2. Example of typical Human MBL standard curve in Assay Diluent B (used for cell culture supernatants). The standard curve was prepared as described in Section 10.
16. **TYPICAL SAMPLE VALUES**

**SENSITIVITY –**
The minimum detectable dose of MBL is 0.03 ng/mL.

**RECOVERY –**
Recovery was determined by spiking various levels of Human MBL into Human serum, plasma and cell culture media. Mean recoveries are as follows:

<table>
<thead>
<tr>
<th>Sample Type</th>
<th>Average % Recovery</th>
<th>Range (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum</td>
<td>122.8</td>
<td>103-147</td>
</tr>
<tr>
<td>Plasma</td>
<td>121.1</td>
<td>98-145</td>
</tr>
<tr>
<td>Cell culture media</td>
<td>103.1</td>
<td>91-116</td>
</tr>
</tbody>
</table>

**LINEARITY OF DILUTION -**

<table>
<thead>
<tr>
<th>Serum Dilution</th>
<th>Average % Expected Value</th>
<th>Range (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1:2</td>
<td>120.4</td>
<td>118-128</td>
</tr>
<tr>
<td>1:4</td>
<td>110.5</td>
<td>103-119</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Plasma Dilution</th>
<th>Average % Expected Value</th>
<th>Range (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1:2</td>
<td>118.8</td>
<td>113-123</td>
</tr>
<tr>
<td>1:4</td>
<td>119.8</td>
<td>109-129</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Cell Culture Media Dilution</th>
<th>Average % Expected Value</th>
<th>Range (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1:2</td>
<td>119.3</td>
<td>111-127</td>
</tr>
<tr>
<td>1:4</td>
<td>120.3</td>
<td>113-128</td>
</tr>
</tbody>
</table>
DATA ANALYSIS

PRECISION –

<table>
<thead>
<tr>
<th></th>
<th>Intra-Assay</th>
<th>Inter-Assay</th>
</tr>
</thead>
<tbody>
<tr>
<td>%CV</td>
<td>&lt;10</td>
<td>&lt;12</td>
</tr>
</tbody>
</table>

17. ASSAY SPECIFICITY

The antibodies used within this ELISA kit detect Human MBL.

The reaction of these antibodies with other species has not been tested.

Please contact our Scientific Support team for more information.
### 18. Troubleshooting

<table>
<thead>
<tr>
<th>Problem</th>
<th>Cause</th>
<th>Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td>Poor standard curve</td>
<td>Inaccurate pipetting</td>
<td>Check pipette performance.</td>
</tr>
<tr>
<td></td>
<td>Improper standards dilution</td>
<td>Prior to opening, briefly spin the stock standard tube and dissolve the powder thoroughly by gentle mixing</td>
</tr>
<tr>
<td>Low Signal</td>
<td>Incubation times too brief</td>
<td>Ensure sufficient incubation time; change to overnight standard/sample incubation</td>
</tr>
<tr>
<td></td>
<td>Inadequate reagent volumes or improper dilution</td>
<td>Check pipette performance and ensure correct preparation</td>
</tr>
<tr>
<td>High %CV</td>
<td>Inaccurate pipetting</td>
<td>Check pipette performance</td>
</tr>
<tr>
<td>High background</td>
<td>Plate is insufficiently washed</td>
<td>Review manual for proper wash technique. If using a plate washer, check all ports for obstructions</td>
</tr>
<tr>
<td></td>
<td>Contaminated wash buffer</td>
<td>Prepare fresh wash buffer</td>
</tr>
<tr>
<td>Low sensitivity</td>
<td>Improper storage of the ELISA kit</td>
<td>Store the reconstituted protein at -80°C, all other assay components 4°C. Keep substrate solution protected from light.</td>
</tr>
<tr>
<td></td>
<td>Stop solution</td>
<td>Stop solution should be added to each well before measurement.</td>
</tr>
</tbody>
</table>
19. NOTES