

ab197011 – Glutamine Assay Kit (Colorimetric)

For the quantitative measurement of Gln in various fluids and tissues.

For research use only - not intended for diagnostic use.

PLEASE NOTE: With the acquisition of BioVision by Abcam, we have made some changes to component names and packaging to better align with our global standards as we work towards environmental-friendly and efficient growth. You are receiving the same high-quality products as always, with no changes to specifications or protocols.

For overview, typical data and additional information please visit:

<http://www.abcam.com/ab197011>

Storage and Stability

On receipt entire assay kit should be stored at -20°C, protected from light. Briefly centrifuge small vials prior to opening. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Materials Supplied

Item	Quantity	Storage Condition
Developer Solution III/Developer	1 vial	-20°C
Assay Buffer XXXI/Development Buffer	25 mL	-20°C
Enzyme Mix VIII/Development Enzyme Mix (lyophilized)	1 vial	-20°C
Glutamine Standard	1 vial	-20°C
Assay Buffer XXIX/Hydrolysis Buffer	25 mL	-20°C
Hydrolysis Enzyme Mix II/Hydrolysis Enzyme Mix	1 vial	-20°C

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

- 96-well clear plate with flat bottom
- Multi-well spectrophotometer
- 10K Spin Column

Reagent Preparation

- Before using the kit, spin the tubes prior to opening.

Assay Buffer XXIX/Hydrolysis Buffer and Assay Buffer XXXI/Development Buffer: Bring to room temperature before use. Store at -20°C. Stable for two months.

Hydrolysis Enzyme Mix II/Hydrolysis Enzyme Mix: Reconstitute with 220 µl Assay Buffer XXIX/Hydrolysis Buffer to make the stock solution. Pipette gently to dissolve. Store at -20°C. Keep on ice while in use. Stable for two months.

Enzyme Mix VIII/Development Enzyme Mix: Reconstitute with 220 µl Assay Buffer XXXI/Development Buffer. Pipette gently to dissolve. Aliquot & store at -20°C. Keep on ice while in use. Stable for two months.

Development Solution III/Developer: Reconstitute with 220 µl Assay Buffer XXXI/Development Buffer. Pipette gently to dissolve. Aliquot & store at -20°C. Keep on ice while in use. Stable for two months.

Gln Standard: Reconstitute with 100 µl ddH₂O to generate 10 mM solution. Store at -20°C. Stable for two months.

Assay Protocol

Sample Preparation:

1. Centrifuge biological fluids at 10,000 X g for 5 min at 4°C. Collect the supernatant & add 1-40 µl into desired well(s) in a 96-well plate.
2. For mammalian tissues, homogenize ~10-20 mg of tissue on ice using 10x (v/w) Assay Buffer XXIX/Hydrolysis Buffer. Centrifuge the homogenate at 10,000 X g, 10 min at 4°C. Collect the supernatant & add 1-40 µl into desired well(s) in a 96-well plate. Adjust the volume to 40 µl/well with ddH₂O.

Δ Notes:

- a. Glutamine concentrations can vary over a wide range depending on the sample. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the Standard Curve range.
- b. Glutamate in the sample will contribute to the background signal. If high glutamate levels are predicted in the sample, prepare parallel sample well(s) as sample background control(s).
- c. For samples having high protein content, we recommend deproteinizing the samples (tissue lysate or biological fluids) using 10K Spin Column. Add sample to the spin column, centrifuge at 10,000 X g for 10 min at 4°C. Collect the filtrate.
- d. Endogenous compounds may interfere with the assay. To ensure accurate determination of Gln in the test samples or for samples having low concentration of Gln, we recommend spiking samples with a known amount of Gln Standard (6 nmol).

Standard Curve Preparation:

1. Dilute Gln Standard to 1 mM by adding 10 µl of 10 mM Gln Standard to 90 µl of ddH₂O.
2. Add 0, 2, 4, 6, 8 and 10 µl of Gln Standard into series of wells in a 96-well plate to generate 0, 2, 4, 6, 8 & 10 nmol/well of Gln Standard.
3. Adjust the volume to 40 µl/well with ddH₂O.

Hydrolysis Mix:

1. Add 2 µl Hydrolysis Enzyme Mix II/Hydrolysis Enzyme mix to the Standard and Sample wells as follows:

Item	Standard/Sample	* Sample Background
Hydrolysis Enzyme Mix II/Hydrolysis Enzyme Mix	2 µl	-
Assay Buffer XXIX/Hydrolysis Buffer	8 µl	10 µl

2. Mix well. Adjust the volume to 50 µl/well with ddH₂O if necessary. Incubate for 30 min at 37 °C.

* For samples having high glutamate levels, add 10 µl of Assay Buffer XXIX/Hydrolysis Buffer to sample background control well(s). Adjust the volume to 50 µl/well with ddH₂O & incubate for 30 min at 37 °C.

Reaction Mix:

- Mix enough reagents for the number of assays to be performed. For each well, prepare 50 µl Reaction Mix containing:

Item	Reaction Mix
Assay Buffer XXXI/Development Buffer	46 µl
Enzyme Mix VIII/Development Enzyme Mix	2 µl
Development Solution III/Developer	2 µl

- Mix well. Add 50 µl of the Reaction Mix to each well containing Standards, samples and Background Control(s). Mix well.

Measurement

Incubate at 37°C for 60 min, protected from light. Measure absorbance (OD 450 nm) in a plate reader.

Calculation

- Subtract 0 Gln Standard reading from all readings. Plot the Gln Standard Curve.
- If sample Background Control reading is significant, then subtract sample Background Control reading from sample reading.
- Apply the corrected OD to the Gln Standard Curve to get B nmol of Gln in the sample well.

$$\text{Sample Gln concentration (C)} = \frac{B}{V} \times D \text{ nmol/}\mu\text{l or mM}$$

Where: **B** = Amount of Gln in the sample well from Standard Curve (nmol)

V = Sample volume added into the reaction well (µL)

D = Sample dilution factor

Δ Note: For spiked samples, correct for any sample interference by using following equation:

$$\text{Gln amount in spiked sample well (B)} = \left(\frac{OD \text{ sample (corrected)}}{(OD \text{ sample} + Gln \text{ Std (corrected)}) - (OD \text{ sample (corrected)})} \right) \times Gln \text{ Spike (nmol)}$$

- Gln concentration can also be expressed as nmol/mg of protein or nmol/mg of creatinine in case of urine.
- Glutamine molecular weight: 146.1 g/mol

Technical Support

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