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ab223590 Mouse Beta-2- Microglobulin SimpleStep ELISA[®] Kit

For the quantitative measurement of Beta-2-Microglobulin in mouse serum, plasma, urine and cell culture supernatant samples.

This product is for research use only and is not intended for diagnostic use.

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1. Overview

Beta-2-Microglobulin *in vitro* SimpleStep ELISA® (Enzyme-Linked Immunosorbent Assay) kit is designed for the quantitative measurement of Beta-2-Microglobulin protein in mouse serum, plasma, urine and cell culture supernatant samples.

The SimpleStep ELISA® employs an affinity tag labeled capture antibody and a reporter conjugated detector antibody which immunocapture the sample analyte in solution. This entire complex (capture antibody/analyte/detector antibody) is in turn immobilized via immunoaffinity of an anti-tag antibody coating the well. To perform the assay, samples or standards are added to the wells, followed by the antibody mix. After incubation, the wells are washed to remove unbound material. TMB substrate is added and during incubation is catalyzed by HRP, generating blue coloration. This reaction is then stopped by addition of Stop Solution completing any color change from blue to yellow. Signal is generated proportionally to the amount of bound analyte and the intensity is measured at 450 nm. Optionally, instead of the endpoint reading, development of TMB can be recorded kinetically at 600 nm.

Beta-2-Microglobulin is the light chain of the class I major histocompatibility complex (MHC); it forms heterodimers with classical (HLA-A, -B, -C) and non-classical (HLA-E, -F, -G) class I MHC heavy chains and with class I non-MHC heavy chains (CD1). Beta-2-Microglobulin is involved in the presentation of peptide antigens to the immune system. Beta-2-Microglobulin can dissociate from the MHC and circulate in complex with alpha 2-Macroglobulin.

2. Protocol Summary

Prepare all reagents, samples, and standards as instructed



Add 50 μ L standard or sample to appropriate wells



Add 50 μ L Antibody Cocktail to all wells



Incubate at room temperature for 1 hour



Aspirate and wash each well three times with 350 μ L 1X Wash Buffer
PT



Add 100 μ L TMB Substrate to each well and incubate for 10 minutes.



Add 100 μ L Stop Solution and read OD at 450 nm

3. Precautions

Please read these instructions carefully prior to beginning the assay.

- All kit components have been formulated and quality control tested to function successfully as a kit.
- We understand that, occasionally, experimental protocols might need to be modified to meet unique experimental circumstances. However, we cannot guarantee the performance of the product outside the conditions detailed in this protocol booklet.
- Reagents should be treated as possible mutagens and should be handled with care and disposed of properly. Please review the Safety Datasheet (SDS) provided with the product for information on the specific components.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- All biological materials should be treated as potentially hazardous and handled as such. They should be disposed of in accordance with established safety procedures.

4. Storage and Stability

Store kit at +4°C immediately upon receipt. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Refer to list of materials supplied for storage conditions of individual components.

5. Limitations

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors. Kits are QC tested as a set of components and performance cannot be guaranteed if utilized separately or substituted.

6. Materials Supplied

| Item | Quantity | Storage Condition |
|--|----------|-------------------|
| Mouse Beta-2-Microglobulin Capture Antibody 10X | 600 µL | +4°C |
| Mouse Beta-2-Microglobulin Detector Antibody 10X | 600 µL | +4°C |
| Mouse Beta-2-Microglobulin Lyophilized Recombinant Protein | 2 Vials | +4°C |
| Antibody Diluent 5BR | 6 mL | +4°C |
| Wash Buffer PT 10X | 20 mL | +4°C |
| Cell Extraction Buffer PTR 5X | 10 mL | +4°C |
| Cell Extraction Enhancer Solution 50X | 1 mL | +4°C |
| Denaturant | 500 µL | +4°C |
| TMB Substrate | 12 mL | +4°C |
| Stop Solution | 12 mL | +4°C |
| Sample Diluent NS | 50 mL | +4°C |
| Pre-coated microplate (12 x 8 well strips) | 96 Wells | +4°C |
| Plate Seal | 1 | +4°C |

7. Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Microplate reader capable of measuring absorbance at 450 or 600 nm.
- Method for determining protein concentration (BCA assay recommended).
- Deionized water.
- Multi- and single-channel pipettes.
- Tubes for standard dilution.
- Plate shaker for all incubation steps.
- Optional: Phenylmethylsulfonyl Fluoride (PMSF) (or other protease inhibitors).

8. Technical Hints

- Samples generating values higher than the highest standard should be further diluted in the appropriate sample dilution buffers.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Complete removal of all solutions and buffers during wash steps is necessary to minimize background.
- As a guide, typical ranges of sample concentration for commonly used sample types are shown below in Sample Preparation (section 11).
- All samples should be mixed thoroughly and gently.
- Avoid multiple freeze/thaw of samples.
- Incubate ELISA plates on a plate shaker during all incubation steps.
- When generating positive control samples, it is advisable to change pipette tips after each step.

- The provided Antibody Diluents and Sample Diluents contain protease inhibitor aprotinin. Additional protease inhibitors can be added if required.
- The provided Cell Extraction Buffer 5X contains phosphatase inhibitors and protease inhibitor aprotinin. Additional protease inhibitors can be added if required.
- The provided Cell Extraction Enhancer Solution 50X may precipitate when stored at + 4°C. To dissolve, warm briefly at +37°C and mix gently. The Cell Extraction Enhancer Solution 50X can be stored at room temperature to avoid precipitation.
- **To avoid high background always add samples or standards to the well before the addition of the antibody cocktail.**
- **This kit is sold based on number of tests. A 'test' simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions.**

9. Reagent Preparation

- Equilibrate all reagents to room temperature (18-25°C) prior to use. The kit contains enough reagents for 96 wells. The sample volumes below are sufficient for 48 wells (6 x 8-well strips); adjust volumes as needed for the number of strips in your experiment.
- Prepare only as much reagent as is needed on the day of the experiment. Capture and Detector Antibodies have only been tested for stability in the provided 10X formulations.

9.1 Sample Diluent NS + Enhancer:

Prepare Sample Diluent NS + Enhancer by diluting 50X Cell Extraction Enhancer Solution to 1X with Sample Diluent NS. To make 10 mL Sample Diluent NS + Enhancer combine 9.8 mL Sample Diluent NS and 200 µL Cell Extraction Enhancer Solution 50X. Mix thoroughly and gently. If required protease inhibitors can be added.

9.2 1X Wash Buffer PT:

Prepare 1X Wash Buffer PT by diluting Wash Buffer PT 10X with deionized water. To make 50 mL 1X Wash Buffer PT combine 5 mL Wash Buffer PT 10X with 45 mL deionized water. Mix thoroughly and gently.

9.3 Antibody Cocktail:

Prepare Antibody Cocktail by diluting the capture and detector antibodies in Antibody Diluent 5BR. To make 3 mL of the Antibody Cocktail combine 300 µL 10X Capture Antibody and 300 µL 10X Detector Antibody with 2.4 mL Antibody Diluent 5BR. Mix thoroughly and gently.

10. Standard Preparation

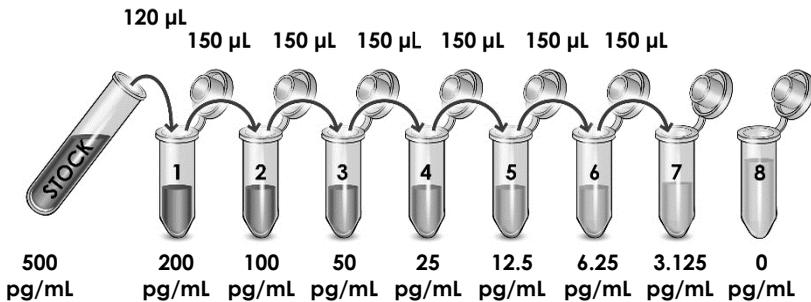
- Always prepare a fresh set of standards for every use.
- Discard working standard dilutions after use as they do not store well.
- The following section describes the preparation of a standard curve for duplicate measurements (recommended).

10.1 Reconstitute the Beta-2-Microglobulin lyophilized standard sample by adding 500 μL Sample Diluent NS + Enhancer. Mix thoroughly and gently. Hold at room temperature for 10 minutes and mix gently. This is the 500 pg/mL **Stock Standard** Solution.

10.2 Label eight tubes, Standards 1– 8.

10.3 Add 180 μL Sample Diluent NS + Enhancer into tube number 1 and 150 μL of Sample Diluent NS + Enhancer into numbers 2-8.

10.4 Use the Stock Standard to prepare the following dilution series. Standard #8 contains no protein and is the Blank control:



11. Sample Preparation

| Typical Sample Dynamic Range | |
|--|-------------------------|
| Sample Type | Range |
| Serum | 1:6,400,000 – 1:100,000 |
| Plasma - Citrate | 1:6,400,000 – 1:100,000 |
| Plasma - Heparin | 1:1,600,000 – 1:100,000 |
| Plasma - EDTA | 1:3,200,000 – 1:100,000 |
| Urine | 1:1600 – 1:100 |
| Lung Tissue Culture Supernatant (6 Days) | 1:32,000 – 1:5,000 |

11.1 Plasma:

Collect plasma using citrate, EDTA or heparin. Centrifuge samples at 2,000 x g for 10 minutes. Store un-diluted plasma samples at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

- 11.1.1 Dilute plasma samples 1:100 in Sample Diluent NS + Enhancer. Mix thoroughly and gently.
- 11.1.2 To 8 volume of plasma add 1 volume of Denaturant. Mix thoroughly and gently.
- 11.1.3 Incubate samples at room temperature for 10 minutes.
- 11.1.4 Dilute samples 40-fold in Sample Diluent NS. Mix thoroughly and gently.
- 11.1.5 Dilute samples as needed to a final concentration in Sample Diluent NS + Enhancer.
- 11.1.6 Example of this dilution scheme is shown below:

| Tube # | Sample to Dilute | Volume to Dilute (µL) | Volume of Sample Diluent NS + Enhancer (µL) | Volume of Denaturant (µL) | Volume of Sample Diluent NS (µL) | Starting Dilution | Final Dilution |
|--------|-------------------|-----------------------|---|---------------------------|----------------------------------|-------------------|----------------|
| 1 | Neat Serum/Plasma | 5 | 495 | 0 | 0 | Neat | 1:100 |
| 2 | Tube #1 | 88.89 | 0 | 11.11 | 0 | 1:100 | 1:112.5 |
| 3 | Tube #2 | 5 | 0 | 0 | 195 | 1:112.5 | 1:4500 |
| 4 | Tube #3 | 9 | 191 | 0 | 0 | 1:4500 | 1:100,000 |

11.2 Serum:

Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 2,000 x g for 10 minutes and collect serum. Store un-diluted serum at -20°C or below. Avoid repeated freeze-thaw cycles.

- 11.2.1 Dilute serum samples 1:100 in Sample Diluent NS + Enhancer. Mix thoroughly and gently.
- 11.2.2 To 8 volume of serum add 1 volume of Denaturant. Mix thoroughly and gently.
- 11.2.3 Incubate samples at room temperature for 10 minutes.
- 11.2.4 Dilute samples 40-fold in Sample Diluent NS. Mix thoroughly and gently.
- 11.2.5 Dilute samples as needed to a final concentration in Sample Diluent NS + Enhancer.
- 11.2.6 Example of this dilution scheme is shown below:

| Tube # | Sample to Dilute | Volume to Dilute (µL) | Volume of Sample Diluent NS + Enhancer (µL) | Volume of Denaturant (µL) | Volume of Sample Diluent NS (µL) | Starting Dilution | Final Dilution |
|--------|--------------------|-----------------------|---|---------------------------|----------------------------------|-------------------|----------------|
| 1 | Neat Serum/ Plasma | 5 | 495 | 0 | 0 | Neat | 1:100 |
| 2 | Tube #1 | 88.89 | 0 | 11.11 | 0 | 1:100 | 1:112.5 |
| 3 | Tube #2 | 5 | 0 | 0 | 195 | 1:112.5 | 1:4500 |
| 4 | Tube #3 | 9 | 191 | 0 | 0 | 1:4500 | 1:100,000 |

11.3 Cell Culture Supernatants:

Centrifuge cell culture media at 2,000 x g for 10 minutes to remove debris. Collect supernatants. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

- 11.3.1 Dilute cell culture supernatant samples as needed (for example 1:50) in Sample Diluent NS + Enhancer. Mix thoroughly and gently.
- 11.3.2 To 8 volume of cell culture supernatant add 1 volume of Denaturant. Mix thoroughly and gently.
- 11.3.3 Incubate samples at room temperature for 10 minutes.
- 11.3.4 Dilute samples 40-fold in Sample Diluent NS. Mix thoroughly and gently.
- 11.3.5 Dilute samples as needed to a final concentration in Sample Diluent NS + Enhancer.
- 11.3.6 Example of this dilution scheme is shown below:

| Tube # | Sample to Dilute | Volume to Dilute (µL) | Volume of Sample Diluent NS + Enhancer (µL) | Volume of Denaturant (µL) | Volume of Sample Diluent NS (µL) | Starting Dilution | Final Dilution |
|--------|------------------|-----------------------|---|---------------------------|----------------------------------|-------------------|----------------|
| 1 | Neat SN | 5 | 245 | 0 | 0 | Neat | 1:50 |
| 2 | Tube #1 | 88.89 | 0 | 11.11 | 0 | 1:50 | 1:56.25 |
| 3 | Tube #2 | 5 | 0 | 0 | 195 | 1:56.25 | 1:2,250 |
| 4 | Tube #3 | 90 | 110 | 0 | 0 | 1:2250 | 1:5,000 |

11.4 Urine:

Centrifuge urine at 2,000 x g for 10 minutes to remove debris. Collect supernatants. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

11.4.1 To 8 volume of urine add 1 volume of Denaturant. Mix thoroughly and gently.

11.4.2 Incubate samples at room temperature for 10 minutes.

11.4.3 Dilute samples 40-fold in Sample Diluent NS. Mix thoroughly and gently.

11.4.4 Dilute samples as needed to a final concentration in Sample Diluent NS + Enhancer. Example of this dilution scheme is shown below:

| Tube # | Sample to Dilute | Volume to Dilute (µL) | Volume of Sample Diluent NS + Enhancer (µL) | Volume of Denaturant (µL) | Volume of Sample Diluent NS (µL) | Starting Dilution | Final Dilution |
|--------|------------------|-----------------------|---|---------------------------|----------------------------------|-------------------|----------------|
| 1 | Neat Urine | 88.89 | 0 | 11.11 | 0 | Neat | 88.89% |
| 2 | Tube #1 | 5 | 0 | 0 | 195 | 88.89% | 2.2225% |
| 3 | Tube #2 | 90 | 110 | 0 | 0 | 2.2225% | 1% |

12. Plate Preparation

- The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents.
- Unused plate strips should be immediately returned to the foil pouch containing the desiccant pack, resealed and stored at 4°C.
- For each assay performed, a minimum of two wells must be used as the zero control.
- For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates).
- Differences in well absorbance or “edge effects” have not been observed with this assay.

13. Assay Procedure

- Equilibrate all materials and prepared reagents to room temperature prior to use.
 - We recommend that you assay all standards, controls and samples in duplicate.
 - Prepare all reagents, working standards, and samples as directed in the previous sections.
- 13.1** Prepare all reagents, working standards, and samples as directed in the previous sections.
 - 13.2** Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, reseal and return to 4°C storage.
 - 13.3** Add 50 µL of all sample or standard to appropriate wells.
 - 13.4** Add 50 µL of the Antibody Cocktail to each well.
 - 13.5** Seal the plate and incubate for 1 hour at room temperature on a plate shaker set to 400 rpm.
 - 13.6** Wash each well with 3 x 350 µL 1X Wash Buffer PT. Wash by aspirating or decanting from wells then dispensing 350 µL 1X Wash Buffer PT into each well. Complete removal of liquid at each step is essential for good performance. After the last wash invert the plate and blot it against clean paper towels to remove excess liquid.
 - 13.7** Add 100 µL of TMB Substrate to each well and incubate for 10 minutes in the dark on a plate shaker set to 400 rpm.
 - 13.8** Add 100 µL of Stop Solution to each well. Shake plate on a plate shaker for 1 minute to mix. Record the OD at 450 nm. This is an endpoint reading.
 - 13.9** Alternative to 13.7 – 13.8: Instead of the endpoint reading at 450 nm, record the development of TMB Substrate kinetically. Immediately after addition of TMB Development Solution begin recording the blue color development with elapsed time in the microplate reader prepared with the following settings:

| Mode | Kinetic |
|-------------|------------------------|
| Wavelength: | 600 nm |
| Time: | up to 15 min |
| Interval: | 20 sec - 1 min |
| Shaking: | Shake between readings |

Δ Note: that an endpoint reading can also be recorded at the completion of the kinetic read by adding 100 μ L Stop Solution to each well and recording the OD at 450 nm.

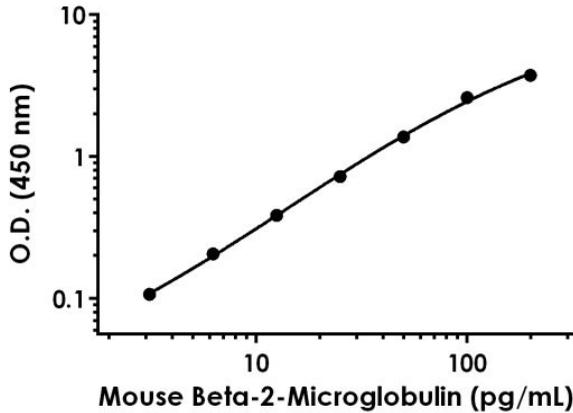
13.10 Analyze the data as described below.

14. Calculations

- 14.1 Calculate the average absorbance value for the blank control (zero) standards. Subtract the average blank control standard absorbance value from all other absorbance values.
- 14.2 **Create a standard curve** by plotting the average blank control subtracted absorbance value for each standard concentration (y-axis) against the target protein concentration (x-axis) of the standard. Use graphing software to draw the best smooth curve through these points to construct the standard curve.
 - Δ **Note:** Most microplate reader software or graphing software will plot these values and fit a curve to the data. A four parameter curve fit (4PL) is often the best choice; however, other algorithms (e.g. linear, semi-log, log/log, 4 parameter logistic) can also be tested to determine if it provides a better curve fit to the standard values.
- 14.3 Determine the concentration of the target protein in the sample by interpolating the blank control subtracted **absorbance values against the standard curve**. Multiply the resulting value by the appropriate sample dilution factor, if used, to obtain the concentration of target protein in the sample.
- 14.4 Samples generating absorbance values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at an absorbance values less than that of the lowest standard should be retested in a less dilute form.

15. Typical Data

Typical standard curve – data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.



| Standard Curve Measurements | | | |
|-----------------------------|------------|-------|----------|
| Concentration (pg/mL) | O.D 450 nm | | Mean O.D |
| | 1 | 2 | |
| 0 | 0.060 | 0.066 | 0.063 |
| 3.125 | 0.168 | 0.171 | 0.170 |
| 6.25 | 0.273 | 0.264 | 0.269 |
| 12.5 | 0.447 | 0.450 | 0.448 |
| 25 | 0.805 | 0.765 | 0.785 |
| 50 | 1.421 | 1.453 | 1.437 |
| 100 | 2.657 | 2.675 | 2.666 |
| 200 | 3.834 | 3.797 | 3.815 |

Figure 1. Example of mouse Beta-2-Microglobulin standard curve in Sample Diluent NS + Enhancer. The Beta-2-Microglobulin standard curve was prepared as described in Section 10. Raw data values are shown in the table. Background-subtracted data values (mean +/- SD) are graphed.

16. Typical Sample Values

SENSITIVITY –

The calculated minimal detectable dose (MDD) is 1.165 pg/mL. The MDD was determined by calculating the mean of zero standard replicates (n=25) and adding 2 standard deviations then extrapolating the corresponding concentration.

RECOVERY –

Three concentrations of Beta-2-Microglobulin were spiked in duplicate to the indicated biological matrix to evaluate signal recovery in the working range of the assay.

| Sample Type | Average % Recovery | Range (%) |
|----------------------------|--------------------|-----------|
| 1:400,000 Serum | 102 | 97 – 106 |
| 1:400,000 Plasma – Citrate | 102 | 101 – 103 |
| 1:400,000 Plasma – Heparin | 93 | 82 – 103 |
| 1:400,000 Plasma – EDTA | 103 | 101 – 107 |
| 1:1,000 Urine | 98 | 95 – 100 |
| 1:10,000 Lung Supernatant | 114 | 110 – 118 |

Linearity of Dilution

Linearity of dilution is determined based on interpolated values from the standard curve. Linearity of dilution defines a sample concentration interval in which interpolated target concentrations are directly proportional to sample dilution.

Native Beta-2-Microglobulin was measured in the following biological samples in a 2-fold dilution series. Sample dilutions are made in Sample Diluent NS + Enhancer.

| Dilution Factor | Interpolated value | 1:100,000 Mouse Serum | 1:100,000 Mouse Plasma (Citrate) | 1:100,000 Mouse Plasma (Heparin) | 1:100,000 Mouse Plasma (EDTA) |
|-----------------|-------------------------|-----------------------|----------------------------------|----------------------------------|-------------------------------|
| Undiluted | pg/mL | 187.9 | 159.5 | 191.0 | 165.8 |
| | % Expected value | 100 | 100 | 100 | 100 |
| 2 | pg/mL | 100.8 | 73.58 | 98.39 | 92.24 |
| | % Expected value | 107 | 92 | 103 | 111 |
| 4 | pg/mL | 48.53 | 36.40 | 47.15 | 38.46 |
| | % Expected value | 103 | 91 | 99 | 93 |
| 8 | pg/mL | 24.51 | 18.64 | 24.83 | 18.95 |
| | % Expected value | 104 | 93 | 104 | 91 |
| 16 | pg/mL | 12.14 | 9.759 | 10.14 | 8.621 |
| | % Expected value | 103 | 98 | 85 | 83 |

| Dilution Factor | Interpolated value | 1:100 Mouse Urine | 1:5,000 Mouse Lung Supernatant |
|-----------------|-------------------------|-------------------|--------------------------------|
| Undiluted | pg/mL | 166.2 | 95.14 |
| | % Expected value | 100 | 100 |
| 2 | pg/mL | 89.76 | 41.55 |
| | % Expected value | 108 | 87 |
| 4 | pg/mL | 43.25 | 21.49 |
| | % Expected value | 104 | 90 |
| 8 | pg/mL | 23.16 | 10.88 |
| | % Expected value | 111 | 91 |
| 16 | pg/mL | 11.98 | 5.592 |
| | % Expected value | 115 | 94 |

PRECISION –

Mean coefficient of variations of interpolated values from three concentrations of mouse serum within the working range of the assay.

| | Intra-Assay | Inter-Assay |
|-------|-------------|-------------|
| n = | 5 | 3 |
| CV(%) | 2.8 | 3.2 |

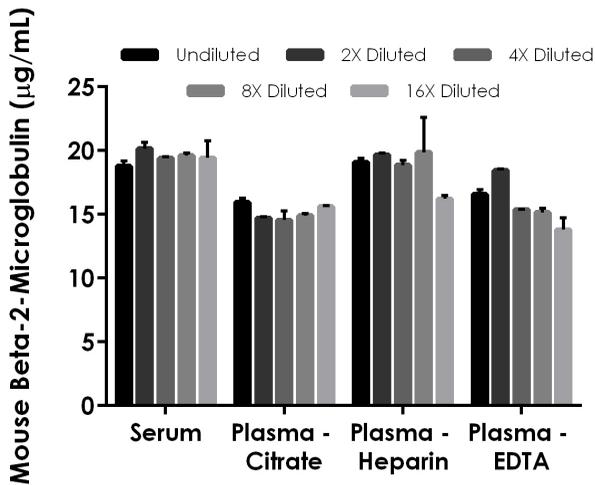


Figure 2. Interpolated concentrations of native Beta-2-Microglobulin in mouse serum, and plasma samples. The concentrations of Beta-2-Microglobulin were measured in duplicates, interpolated from the Beta-2-Microglobulin standard curves and corrected for sample dilution. Undiluted samples are as follows: serum 1:100,000, plasma (citrate) 1:100,000, plasma (heparin) 1:100,000, and plasma (EDTA) 1:100,000. The interpolated dilution factor corrected values are plotted (mean +/- SD, n=2). The mean Beta-2-Microglobulin concentration was determined to be 19.48 µg/mL in neat serum, 15.15 µg/mL in neat plasma (citrate), 18.74 µg/mL in neat plasma (heparin), and 15.87 µg/mL in neat plasma (EDTA).

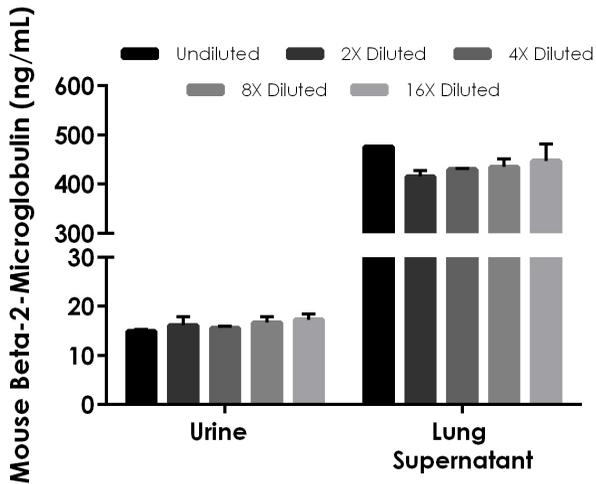


Figure 3. Interpolated concentrations of native Beta-2-Microglobulin in mouse urine and lung tissue culture supernatant samples. The concentrations of Beta-2-Microglobulin were measured in duplicates, interpolated from the Beta-2-Microglobulin standard curves and corrected for sample dilution. Undiluted samples are as follows: urine 1:100, lung supernatant 1:5,000. The interpolated dilution factor corrected values are plotted (mean +/- SD, n=2). The mean Beta-2-Microglobulin concentration was determined to be 16.12 ng/mL in neat urine, 440.7 ng/mL in neat lung tissue culture supernatant.

17. Assay Specificity

This kit recognizes both native and recombinant mouse Beta-2-Microglobulin protein in serum, plasma, urine, and cell culture supernatant samples only.

Cell and tissue extract samples have not been tested with this kit.

INTERFERENCE

Recombinant mouse CD1D recombinant protein was prepared at 50 ng/mL and 1 ng/mL and tested for interference. No interference with CD1D was observed.

18. Species Reactivity

This kit recognizes mouse Beta-2-Microglobulin protein.

This kit is not compatible with rat samples.

Other species reactivity was not determined.

Please contact our Technical Support team for more information.

19. Troubleshooting

| Problem | Reason | Solution |
|-------------------------------|--|--|
| Poor standard curve | Inaccurate Pipetting | Check pipettes |
| | Improper standard dilution | Prior to opening, briefly spin the stock standard tube and dissolve the powder thoroughly by gentle mixing |
| Low Signal | Incubation times too brief | Ensure sufficient incubation times; increase to 2 or 3 hour standard/sample incubation |
| | Inadequate reagent volumes or improper dilution | Check pipettes and ensure correct preparation |
| | Incubation times with TMB too brief | Ensure sufficient incubation time until blue color develops prior addition of Stop solution |
| Large CV | Plate is insufficiently washed | Review manual for proper wash technique. If using a plate washer, check all ports for obstructions. |
| | Contaminated wash buffer | Prepare fresh wash buffer |
| Low sensitivity | Improper storage of the ELISA kit | Store your reconstituted standards at -80°C, all other assay components 4°C. Keep TMB substrate solution protected from light. |
| Precipitate in Diluent | Precipitation and/or coagulation of components within the Diluent. | Precipitate can be removed by gently warming the Diluent to 37°C. |

20. Notes

Technical Support

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