

Version 1b Last updated 4 May 2021

# ab237656 anti-Infliximab ELISA Kit

For the measurement of the antibody against Infliximab in human serum and plasma.

This product is for research use only and is not intended for diagnostic use.

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# 1. Overview

Anti-Infliximab ELISA Kit (ab237656) is designed to quantify/measure the antibody against Infliximab with high specificity and sensitivity in biological matrices.

Infliximab is a therapeutic chimeric monoclonal antibody against tumor necrosis factor (TNF) and is used to treat rheumatic arthritis, intestinal disorders, dermatological diseases and cancer. Infliximab drug inhibits the action of TNF and reduces the inflammation and subsequently improves the patient's health. Drug level quantification can be important to adapt patient prescription or to switch to an alternative TNF inhibitor drug. Infliximab is used for the treatment of psoriasis, Crohn's disease, ankylosing spondylitis, psoriatic arthritis, rheumatoid arthritis, ulcerative colitis. However, some patients develop unwanted immunogenicity, which leads to production of anti-drug antibodies (ADAs) inactivating the therapeutic effects of the treatment and, in rare cases, inducing adverse effects.

## 2. Protocol Summary

Prepare all reagents, samples, and standards as instructed



Add 100  $\mu$ L standard, controls, diluted sample and confirmation test mixture to appropriate wells. Cover and incubate for 60 minutes at room temperature



Discard incubation solution and wash plate 3 times with 300  $\mu$ L diluted Wash Buffer



Add 100  $\mu$ L peroxidase conjugate to each well. Cover and incubate for 60 minutes at room temperature



Discard the solution and wash plate 3 times with 300  $\mu$ L diluted Wash Buffer



Add 100  $\mu$ L TMB Substrate and incubate the plate in the dark at room temperature for 20 minutes.



Add 100  $\mu$ L Stop Solution and read OD at 450 nm within 20 minutes.

### 3. Precautions

Please read these instructions carefully prior to beginning the assay.

- Reagents should be treated as possible mutagens and should be handled with care and disposed of properly. Please review the Safety Datasheet (SDS) provided with the product for information on the specific components.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- For general guidelines, precautions, limitations on the use of our assay kits and general assay troubleshooting tips, particularly for first time users, please consult our guide: [www.abcam.com/assaykitguidelines](http://www.abcam.com/assaykitguidelines)
- All biological materials should be treated as potentially hazardous and handled as such. They should be disposed of in accordance with established safety procedures.

### 4. Storage and Stability

Store kit at +4°C immediately upon receipt. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Refer to list of materials supplied for storage conditions of individual components.

### 5. Limitations

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors.

### 6. Materials Supplied

Item	Quantity	Storage
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		<b>condition</b>
Micro ELISA Plate	1 unit	+4°C
Infliximab Standard S1	1 mL	+4°C
Infliximab Standard S2	1 mL	+4°C
Infliximab Standard S3	1 mL	+4°C
Infliximab Standard S4	1 mL	+4°C
Infliximab Standard S5	1 mL	+4°C
Infliximab Standard S6	1 mL	+4°C
Infliximab Standard S7	1 mL	+4°C
Confirmation Reagent	12 mL	+4°C
Assay Buffer	50 mL	+4°C
Peroxidase Conjugate	12 mL	+4°C
TMB Substrate	12 mL	+4°C
Stop Solution	12 mL	+4°C
Wash Buffer (20X)	50 mL	+4°C
Plate sealers	2 units	+4°C

## 7. Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Microplate reader capable of measuring absorbance at OD 450 nm
- Deionized water.
- Multi- and single-channel pipettes.
- Tubes for sample dilution.
- Plate shaker for all incubation steps.
- Absorbent paper

## 8. Technical Hints

- Samples generating values higher than the highest standard should be further diluted.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Complete removal of all solutions and buffers during wash steps is necessary to minimize background.
- All samples should be mixed thoroughly and gently.
- Avoid multiple freeze/thaw of samples.
- Incubate ELISA plates on a plate shaker during all incubation steps.
- When generating positive control samples, it is advisable to change pipette tips after each step.

## 9. Reagent Preparation

- Equilibrate all reagents to room temperature (18-25°C) prior to use. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
- Prepare only as much reagent as is needed on the day of the experiment.

### 9.1 20X Wash Buffer:

Dilute the 20X Wash Buffer to 1X solution in ddH<sub>2</sub>O (10 mL of Wash Buffer stock to 190 mL of ddH<sub>2</sub>O). Mix the 1X solution thoroughly by vortex manually. The working stock can be stable for 2 weeks after preparation at 4°C.

## 10. Standard and Control Preparation

Standard and controls, S1 – S7, are ready to use, please see table below for concentrations:

Name	S1	S2	S3	S4	S5	S6	S7
Conc. ng/mL	500	250	125	62	0	Low control	High control

Concentration for high and low controls are indicated on vials.



## 11. Sample Preparation

### General sample information:

- We recommend performing several dilutions of your sample to ensure the readings are within the standard value range.
- We recommend that you use fresh samples for the most reproducible assay.

### 11.1 Serum/plasma:

1. Dilute samples at 1:10 (20  $\mu$ L serum/plasma + 180  $\mu$ L ddH<sub>2</sub>O) or 1:100 (5  $\mu$ L serum/plasma + 495  $\mu$ L ddH<sub>2</sub>O).
2. Diluted samples should further be diluted if the concentration of Infliximab is higher than the measuring range.
3. Samples are stable at 4°C for 7 days and -20°C for 6 months. Avoid freeze-and-thaw cycle.

**Δ Note:** The usual precautions for venipuncture should be observed.

## 12. Confirmation test mixture preparation

Mix 20  $\mu$ L undiluted (positive) serum/plasma sample with 180  $\mu$ L of confirmation reagent for 60 minutes in a microtube prior to the test.

## 13. Assay Procedure

- Prepare reagents within 30 minutes before the experiment.
  - Equilibrate all materials and prepared reagents to room temperature 15 minutes prior to use.
  - We recommend that you assay all standards, controls and samples in duplicate.
- 13.1** Add 100  $\mu\text{L}$  of standards, controls, diluted samples and confirmation test mixture into appropriate wells. Cover wells and incubate for 60 minutes at room temperature.
  - 13.2** Discard incubation solution. Wash plate 3 times each with 300  $\mu\text{L}$  of diluted Wash Buffer. Remove excess solution by tapping the inverted plate on a paper towel.
  - 13.3** Add 100  $\mu\text{L}$  of Peroxidase Conjugate into each well. Cover wells with adhesive plate sealer and incubate at room temperature for 60 minutes.
  - 13.4** Discard the solution and wash the wells as step 13.2.
  - 13.5** Add 100  $\mu\text{L}$  of 1X TMB substrate solution and incubate the plate in the dark at room temperature for 20 minutes.
  - 13.6** Add 100  $\mu\text{L}$  of Stop solution to stop the reaction.
  - 13.7** Read the absorbance in a microplate reader set to 450 nm within 20 minutes. (Reference wavelength to 650 nm).

## 14. Calculations

### 14.1 Quantitative Calculation

- 14.1.1 Calculate the average absorbance value for the blank control (0 ng/mL) standards. Subtract the average blank control standard absorbance value from all other absorbance values.
- 14.1.2 **Create a standard curve** by plotting the average blank control subtracted absorbance value for each standard concentration (y axis) against the target protein concentration (x axis) of the standard.
- 14.1.3 Construct a standard curve of difference data using software capable of generating four-parameter logistic (4PL) or point-to-point calculation curve fit.
- 14.1.4 To obtain the exact values of the samples, the concentration determined from the standard curve should be multiplied by the dilution factor.
- 14.1.5 Determine the concentration of the target protein in the sample by interpolating the blank control subtracted **absorbance values against the standard curve**. Multiply the resulting value by the appropriate dilution factor to obtain the concentration of target protein in the sample.
- 14.1.6 Samples generating absorbance values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at an absorbance values less than that of the lowest standard should be retested in a less dilute form.

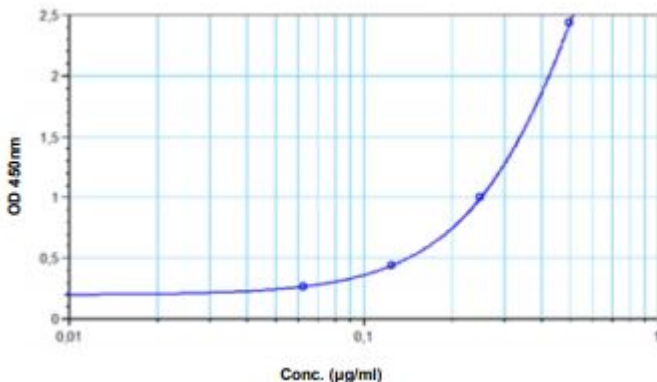
### 14.2 Qualitative Interpretation

- 14.2.1 If "Sample OD<sub>450/650</sub> / Zero Standard (S5) OD<sub>450/650</sub>" is < 3, the sample is NEGATIVE for Antibody to Infliximab (ATR).
- 14.2.2 If "Sample OD<sub>450/650</sub> /Zero Standard (S5) OD<sub>450/650</sub>" is ≥3, the sample is POSITIVE for ATR, and if required samples may be extrapolated for quantitative analysis and confirmation.
- 14.2.3 For the run to be valid, the OD<sub>450/650</sub> nm of High Control should be ≥ 1.000 and the OD<sub>450/650</sub> nm of each Low Control should be <0.200, if not, improper technique or reagent deterioration may be suspected and the run should be repeated.
- 14.2.4 Interpretation of true and false positive: For true positive sample, inhibition should be equal or greater than 25%.

$$\frac{OD \text{ sample} - OD \text{ sample with confirmation reagent}}{OD \text{ sample}} \times 100 = \text{inhibition \%}$$

## 15. Typical Data

Typical standard curve - data provided **for demonstration purposes only**. A new standard curve must be generated for each assay performed.



**Figure 1.** Typical Standard Curve: This standard curve is for demonstration only. A standard curve must be run with each assay.

## 16. Typical Sample Values

**Detection Range:** 62 - 500 ng/mL.

**Sensitivity:** 15 ng/mL.

**Assay Precision:** Intra-Assay: CV < 15%; Inter-Assay: CV < 15%

(CV (%) = SD/mean X 100)

**Cross Reactivity:** Infliximab infusion camouflages/masks the presence of antibody to infliximab (ATI) in serum/plasma samples. Therefore, blood sampling time is critical for detection of ATI. It is convenient to obtain blood sample just before the infusion of infliximab or at least 2 weeks after the infusion of infliximab.

**Recovery rate:** 85 – 115% with normal human serum samples with known concentrations.

## 17. Notes



## Technical Support

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