ab245884
Movat Pentachrome Stain Kit (Modified Russell-Movat)

For the histological visualization of collagen, elastin, muscle, mucin and fibrin in tissue sections.

View Movat Pentachrome Stain Kit (Modified Russell-Movat) datasheet:
www.abcam.com/ab245884
(use www.abcam.cn/ab245884 for China, or www.abcam.co.jp/ab245884 for Japan)

This product is for research use only and is not intended for diagnostic use.
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1. Overview

The Movat Pentachrome Stain Kit (Modified Russel-Movat, ab245884) is intended for use in histological demonstration of collagen, elastin, muscle, mucin and fibrin in tissue sections. This procedure is particularly useful when studying the heart, blood vessels and various vascular diseases.

**Staining Interpretation:**

<table>
<thead>
<tr>
<th>Component</th>
<th>Color</th>
</tr>
</thead>
<tbody>
<tr>
<td>Elastic Fibres</td>
<td>Black to Blue/Black</td>
</tr>
<tr>
<td>Nuclei:</td>
<td>Blue/Black</td>
</tr>
<tr>
<td>Collagen:</td>
<td>Yellow to Red</td>
</tr>
<tr>
<td>Reticular Fibres:</td>
<td>Yellow</td>
</tr>
<tr>
<td>Mucin:</td>
<td>Bright Blue</td>
</tr>
<tr>
<td>Fibrin:</td>
<td>Bright Red</td>
</tr>
<tr>
<td>Muscle:</td>
<td>Red</td>
</tr>
</tbody>
</table>

**Control Tissue:**

Lung, Skin, Colon, Heart or any vascular tissue.
2. Materials Supplied and Storage

Store kit at Room temperature immediately on receipt and check below for storage for individual components. Kit can be stored for 1 year from receipt, if components have not been reconstituted.

Keep away from open flame and refer to the safety datasheet.

<table>
<thead>
<tr>
<th>Item</th>
<th>Quantity</th>
<th>Storage temperature (before prep)</th>
<th>Storage temperature (after prep)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hematoxylin Solution (5%)</td>
<td>250 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Ferric Chloride Solution (10%)</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Lugol’s Iodine Solution</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Ferric Chloride (2%) Differentiating Solution</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Sodium Thiosulfate Solution (5%)</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Acetic Acid Solution (3%)</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Acetic Acid Solution (1%)</td>
<td>250 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Alcian Blue Solution, pH 2.5</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Biebrich Scarlet – Acid Fuchsin Solution</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Phosphotungstic Acid Solution (5%)</td>
<td>250 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
<tr>
<td>Metanil Yellow Solution</td>
<td>125 mL</td>
<td>RT</td>
<td>RT</td>
</tr>
</tbody>
</table>
3. General guidelines, precautions, and troubleshooting

Please observe safe laboratory practice and consult the safety datasheet.

For general guidelines, precautions, limitations on the use of our assay kits and general assay troubleshooting tips, particularly for first time users, please consult our guide: www.abcam.com/assaykitguidelines

For typical data produced using the assay, please see the assay kit datasheet on our website.
4. Reagent Preparation

1. Prepare a working Elastic Stain Solution by mixing 30 ml of Hematoxylin (5%) Solution, 15 ml of Ferric Chloride Solution (10%) and 15 mL of Lugol’s Iodine Solution.

△Note: Mixed solution may be used for 24 hours.
△Note: Lugol’s Iodine Solution will cause staining of all kit vials and labels over time. This does not adversely affect the performance of this product and is merely cosmetic in nature.
△Note: Removal of mercury deposits is not required for tissues that have been fixed in mercury containing fixatives since it will be removed by the staining solution.

5. Assay Procedure

- Equilibrate all materials and prepared reagents to room temperature just prior to use and gently agitate.

1.2 Deparaffinize sections if necessary and hydrate to distilled water.
1.3 Stain tissue section with working Elastic Stain Solution for 20 minutes.
1.4 Rinse in running tap water until no excess stain remains on slide.
1.5 Dip slide in Ferric Chloride (2%) Differentiating Solution 15-20 times and rinse in tap water.
1.6 Check slides microscopically for proper differentiation. Repeat step 4 if required.
1.7 Rinse in 2 changes of distilled water.
1.8 Place slide in Sodium Thiosulfate Solution (5%) and incubate for 1 minute.
1.9 Rinse in tap water for 2 minutes followed by 2 changes in distilled water.
1.10 Place slide in Acetic Acid Solution (3%) and incubate for 2 minutes to equilibrate tissue prior to staining with Alcian Blue Solution, pH 2.5.
1.11 Without rinsing, place slide in Alcian Blue Solution, pH 2.5 and incubate for 25 minutes.
1.12 Rinse in tap water for 2 minutes followed by 2 changes in distilled water.
1.13 Place slide in Biebrich Scarlet – Acid Fuchsin Solution and incubate for 2 minutes.
1.14 Rinse slide in 2 changes of distilled water.
1.15 Place slide in Acetic Acid Solution (1%) for 5-10 seconds with agitation.
1.16 Rinse quickly in distilled water.
1.17 Differentiate slide in 2 changes of Phosphotungstic Acid Solution (5%) for 3-7 minutes each.
1.18 Check slides microscopically for proper differentiation. Collagen should be clear but elastic fibers should still be stained. Repeat step 16 if required.
1.19 Rinse slide briefly in distilled water.
1.20 Dip slide several times (3-5) in Acetic Acid Solution (1%).
1.21 Shake off excess Acetic Acid Solution (1%) and without rinsing apply Metanil Yellow Solution and incubate for 15 minutes.
1.22 Rinse slide in 3 changes of absolute alcohol.
1.23 Clear, and mount in synthetic resin.
6. Notes
Technical Support

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Austria
wissenschaftlicherdienst@abcam.com | 019-288-259

France
supportscientifique@abcam.com | 01.46.94.62.96

Germany
wissenschaftlicherdienst@abcam.com | 030-896-779-154

Spain
soportecientifico@abcam.com | 91-114-65-60

Switzerland
technical@abcam.com

UK, EU and ROW
technical@abcam.com | +44(0)1223-696000

Canada
can.technical@abcam.com | 877-749-8807

US and Latin America
us.technical@abcam.com | 888-772-2226

Asia Pacific
hk.technical@abcam.com | (852) 2603-6823

China
cn.technical@abcam.com | 400 921 0189 | +86 21 2070 0500

Japan
technical@abcam.co.jp | +81-(0)3-6231-0940

Singapore
sg.technical@abcam.com | 800 188-5244

Australia
au.technical@abcam.com | +61-(0)3-8652-1450

New Zealand
nz.technical@abcam.com | +64-(0)9-909-7829